

RESEARCH ARTICLE

Development and Validation of a HPLC Method for Nelfinavir and Tenofovir in Pharmaceutical Dosage Forms

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ABSTRACT

The proposed HPLC method was found to be simple, specific, precise, accurate, rapid and economical for simultaneous estimation of Nelfinavir and Tenofovir in tablet dosage form. The developed method was validated in terms of accuracy, precision, linearity, robustness and ruggedness, and results will be validated statistically according to ICH guidelines. The Sample recoveries in all formulations were in good agreement with their respective label claims. From literature review and solubility analysis initial chromatographic conditions Mobile phase ACN: ortho Phosphate buffer pH 4.0 (70: 30 % v/v) were set (Buffer P^H 2.45 adjusted with Triethylamine), Agilent C18 column (4.6×150 mm) 5µ Column, Flow rate 1.0 ml/min and temperature was ambient, eluent was scanned with PDA detector in system and it showed maximum absorbance at 230 nm. As the methanol content was increased Nelfinavir and Tenofovir got eluted with good peak symmetric properties. The retention times for Nelfinavir and Tenofovir was found to be 2.462 & 3.737 min respectively. System suitability parameters were studied by injecting the standard five times and results were well under the acceptance criteria. Linearity study was carried out between 50% to150 % levels, R² value was found to be as 0.999 by using above method assay of marketed formulation was carried out, 100.7% was present. Full length method was not performed; if it is done this method can be used for routine analysis of Nelfinavir and Tenofovir.

Keywords: HPLC, Nelfinavir, Tenofovir, Ortho Phosphoric Acid Buffer: ACN, Agilent C18.

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1. Introduction

Nelfinavir is a potent HIV-1 protease inhibitor. It is used in combination with other antiviral drugs in the treatment of HIV in both adults and children. Nelfinavir inhibits the HIV viral proteinase enzyme which prevents cleavage of the gag-pol polyprotein, resulting in noninfectious, immature viral particles.



Figure 1 Nelfinavir

IUPAC Name: (3S,4aS,8aS)-N-tert-butyl-2-[(2R, 3R)-2hydroxy-3-[(3-hydroxy-2-methylphenyl)formamido]-4-(phenylsulfanyl)butyl]-decahydroisoquinoline-3carboxamide

Chemical formula: C32H45N3O4SMolecular weight: 567.782

Tenofovir inhibits the activity of HIV reverse transcriptase by competing with the natural substrate deoxyadenosine 5'-triphosphate and, after incorporation into DNA, by DNA chain termination



Figure 2 Tenofovir

IUPAC Name: 4-(1-hydroxy-2-{[6-(4-phenylbutoxy) hexyl] amino} ethyl)-2-(hydroxymethyl)phenol Chemical formula: C₂₅H₃₇NO₄ Molecular weight: 415.5656 g/mol pKa: 2.86

Table 1 List of showingle and standards

2. Methodology

radie.1 List of chemicals and standards used				
S.No	Chemicals	Manufacturer Name	Grade	
1.	Water	Merck	HPLC grade	
2.	Methanol	Merck	HPLC grade	
3.	Acetonitrile	Merck	HPLC grade	
4.	Ortho phosphoric acid	Merck	G.R	
5.	KH ₂ PO ₄	Merck	G.R	
6.	K ₂ HPO ₄	Merck	G.R	
7.	0. 22µ Nylon	Advanced lab	HPLC grade	

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	filter			
8.	0.45µ filter	Millipore	HPLC grade	
	paper	winipore		
9.	Nelfinavir	In House	In- House	
	and Tenofovir	III – House		

Trial-1

Chromatographic conditions Column : Symmetry C18 4.6x150mm, 5µm Mobile phase ratio: ACN: H2O (70:30% v/v) Detection wavelength : 240nm Flow rate: 1ml/min Injection volume : 10µl Run time : 5min Retention time: 1.250 min&1.496 min



Figure 3. Chromatogram showing trial 1 injection

Observation: The trial shows no proper separation of peaks in the chromatogram, so more trials were required for obtaining peaks.

Trial - 2

Chromatographic conditions Column : Inertsil C18 4.6x150mm 5 μ m Mobile phase ratio: Methanol: H2O (70:30% v/v) Detection wavelength: 230 nm Flow rate: 1ml/min Injection volume : 10 μ l Column temperature: Ambient Auto sampler temperature: Ambient Run time: 10.0 min Retention time: 1,536 min & 3,584 mins



Figure 4. Chromatogram showing trial-2 injection

Observation

In this trial both peaks were eluted well, but there was no proper baseline. Need some more trails.

Optimized method:

Chromatographic conditions Column : Agilent column (4.6×150mm)5µ Mobile phase ratio: ACN: Phosphate buffer pH 4.0 (70: 30 % v/v) Detection wavelength: 230 nm Flow rate: 1.0ml/min Injection volume : 10µl Column temperature: Ambient Auto sampler temperature : Ambient Run time: 10min

Retention time: 2.462 & 3.737 mins



Figure 5. Optimized Chromatogram

Observation

The separation was good, peak shape was good, so we conclude that there is no required for reduce the retention times of peaks, so it is taken as final method.

Preparation of phosphate buffer

2.95 grams of KH_2PO_4 and 5.45 grams of K_2HPO_4 was weighed and taken into a 1000ml beaker, dissolved and diluted to 1000ml with HPLC water and pH was adjusted to 3 with ortho phosphoric acid. The resulting solution was sonicated and filtered.

Preparation of mobile phase

Mix a mixture of above buffer 300 ml (30%) and 700 ml of methanol (HPLC grade-70%) and degassed in ultrasonic water bath for 5 minutes. Filter through 0.22 μ filter under vacuum filtration.

Diluents preparation

Mobile phase was used as the diluent.

Preparation of the Nelfinavir and Tenofovir standard and sample solution

Sample solution preparation:

An equivalent tablet power such that 5 mg of Nelfinavir and 2 mg Tenofovir tablet powder were accurately weighed and transferred into a 10 ml clean dry volumetric flask, add about 2ml of diluent and sonicate to dissolve it completely and making volume up to the mark with the same solvent(Stock solution). Further pipette 1ml of the above stock solution into a 100ml volumetric flask and was diluted up to the mark with diluent. (Concentration is 50 ppm for Nelfinavir and 20 ppm for Tenofovir)

Standard solution preparation

5 mg Nelfinavir and 2 mg Tenofovir working standard was accurately weighed and transferred into a 10ml clean dry volumetric flask and add about 2ml of diluent and sonicate to dissolve it completely and make volume up to the mark

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with the same solvent (Stock solution).Further pipette out 1ml of the above stock solution into a 10ml volumetric flask and was diluted up to the mark with diluent. (Concentration is 50 ppm for Nelfinavir and 20 ppm for Tenofovir)

System suitability

5 mg of Nelfinavir and 2mg of Tenofovir working standard was accurately weighed and transferred into a 10ml clean dry volumetric flask and add about 2ml of diluent and sonicate to dissolve it completely and make volume up to the mark with the same solvent (Stock solution).Further pipette out 1ml of Nelfinavir and Tenofovir from the above stock solution into a 10ml volumetric flask and was diluted up to the mark with diluent. (Concentration is 50 ppm for Nelfinavir and 20 ppm for Tenofovir)

3. Results and Discussion



Figure 6. Spectrum showing overlapping spectrum of Nelfinavir and Tenofovir



Figure 7. Chromatogram showing blank preparation (mobile phase)



Figure 8. Assay calculation for Nelfinavir and Tenofovir

	Name	Rt	Area
1	Nelfinavir	2.536	2497318
2	Nelfinavir	2.536	2494274
Mean			2495796
Std.dev			1152.9
%RSD			0.06



Figure 9. Assay calculation for Nelfinavir and Tenofovir

	Name	Rt	Area
1	Tenofovir	3.265	958190
2	Tenofovir	3.265	965083
Mean			961637
Std.dev			38733
%RSD			0.48



Figure 10. Chromatogram showing standard injection



Figure 11. Chromatogram showing sample injection



Figure 12. Calibration curve of Nelfinavir



Figure 13. Calibration curve of Tenofovir

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	Peak name	RT	Area		
1	Tenofovir	3.230	925541		
2	Tenofovir	3.239	923214		
3	Tenofovir	3.246	923881		
4	Tenofovir	3.257	920840		
5	Tenofovir	3.271	926447		
Mean			923984.6		
Std.dev			1948.274		
%RSD			0.210856		

Table 3. Showing	%RSD	results	for	Nelfina	ıvir
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	Peak name	RT	Area
1	Nelfinavir	2.506	2367917
2	Nelfinavir	2.516	2324161
3	Nelfinavir	2.519	2390163
4	Nelfinavir	2.531	2323428
5	Nelfinavir	2.544	2329454
Mean			2347025
Std.dev			27150.26
%RSD			1.156795



Figure 14. Showing results LOD



Figure 15. Showing results LOQ

4. Conclusion

A new method was established for simultaneous estimation of Nelfinavir and Tenofovir by RP-HPLC method. The chromatographic conditions were successfully developed for the separation of Nelfinavir and Tenofovir by using Agilent column (4.6×150 mm) 5μ , flow rate was 1.0 ml/min, mobile phase ratio was (70:30 v/v) ACN: phosphate buffer(KH₂PO₄and K₂HPO₄) phosphate pH 4.0 (pH was adjusted with orthophosphoricacid),detection wavelength was 230 nm. The instrument used was WATERS HPLC Auto Sampler, Separation module 2695, photo diode array detector 996, Empower-software version-2. The retention times were found to be 2.462 & 3.737mins. The % purity of Nelfinavir and Tenofovir was found to be 99.87% and 100.27% respectively. The system suitability parameters for Nelfinavir and Tenofovir such as theoretical plates and tailing factor were found to be 2733, 1.6 and 3500 and 1.4, the resolution was found to be 4.6. The analytical method was validated according to ICH guidelines (ICH, Q2 (R1)).

5. Reference

- [1] Dr. Kealey and P.J Haines, Analytical Chemistry, 1stedition, Bios Publisher, (2002), PP 1-7.
- [2] A.BraithWait and F.J.Smith, Chromatographic Methods, 5thedition, Kluwer Academic Publisher, (1996), PP 1-2.
- [3] Andrea Weston and Phyllisr. Brown, HPLC Principle and Practice, 1st edition, Academic press, (1997), PP 24-37.
- [4] Yuri Kazakevich and Rosario Lobrutto, HPLC for Pharmaceutical Scientists, 1stedition, Wiley Interscience A JohnWiley & Sons, Inc., Publication, (2007), PP 15-23.
- [5] Meyer V.R. Practical High-Performance Liquid Chromatography, 4th Ed. England, John Wiley & Sons Ltd, (2004), PP 7-8.
- [6] Sahajwalla CG a new drug development, vol 141, Marcel Dekker Inc., New York, (2004), PP 421– 426.
- [7] Snyder LR practical HPLC method development, 2nd edition. John Wiley and sons, New York, (1997), PP 180-182.
- [8] Lim SG, Ng TM, Kung N et al. (January 2006). "A double-blind placebo-controlled study of emtricitabine in chronic hepatitis B". Arch. Intern. Med. 166 (1): 49–56.
- [9] Tenofovir". PubChem Public Chemical Database. NCBI. Retrieved 2011-04-10.Jeffries, D. J. (1989).
 "Tenofovir resistant HIV". BMJ (Clinical research ed.) 298 (6681): 1132–1133. PMID 2500164.
- [10] Halde Supriya, Mungantiwar Ashish1 and Chintamaneni Meena2. Journal of Chemical and Pharmaceutical Research, 2012, 4(1):254-259.
- [11] Deepthi Komaroju, G. Nagarjuna Reddy, K. Dhanalakshmi. International Journal of Pharma Research & Review, Oct 2013; 2(10):1-11.
- [12] Nagaraju P.T,K. P. Channabasavaraj, Shantha Kumar P. T. International Journal of ChemTech ResearchCODEN(USA): IJCRGG ISSN : 0974-4290Vol.3, No.1, pp 23-28, Jan-Mar 2011.
- [13] Patel Bhavini N pradeep kumar*1, s.c.dwivedi1, ashok kushnoor2. farmacia, 2012, Vol. 60, 3. Palani Venkatesha, MaheshDaggumatib, Journal of Pharmaceutical Analysis 2012;2(2):152–155.
- [14] Yadavalli Rekha, Yellina Haribabu2, Sheeja Velayudhankutty, Sosamma Cicy Eapen2 and Jane Mary2. Research Journal of Pharmaceutical Science. ISSN 2319 – 555X Vol. 2(4), 10-18, May (2013)
- [15] CH Venkata Reddiah 1, P. Rama Devi 2, K. Mukkanti 3, Srinivasarao Katari. Int.J.Pharm.Phytopharmacol.Res. 2012, 1(5): 247-256.

- [16] Devyani dube and s. p. vyas*. international journal of pharmacy and pharmaceutical sciences.vol 1, issue 2.oct –dec 2009.
- [17] Narendra Devanaboyina*, Anupama Barik, D.Indrani, S. Pooja, S.Vaishnavi, U.Aparna Rajeevi. International Journal of Science Innovations and Discoveries. IJSID, 2012, 2 (1), 170-178.