

Design and development of Ethosomal Gel of Diacerein by using cold method

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ABSTRACT

Trans dermal route offers several potential advantages over conventional routes. These advantages includes avoidance of first pass metabolism, predictable and extended duration of action, minimizing undesirable side effects, utility of short halflife drugs, improving physiological and pharmacological response, avoiding the fluctuation in the blood levels, and most important it provides patient convenience. Recently advancement in liposomes was done and result obtained was "Ethosomal system". Ethosomal system showed topical delivery with higher trans dermal flux and higher skin deposition and became an attractive option as it has several desirable advantages. Diacerein is under investigation for the treatment of Insulin Resistance, Diabetes Mellitus (Type 2), and Diabetes-Related Complications. Diacerein Ethosomes were prepared using the method reported by Touitou et al., (2000) with little modification. Studies were performed on Ethosomes containing 20%, 30%, 40% and 50% w/w ethanol with sonication used for their size reduction. To confirm the presence of vesicular structure, formulations were visualized under microscope at different magnified fields, which showed presence of lipid bilayer as well as spherical structure of vesicles. Using the same microscopic method and special software "particle size analysis", size of vesicle was determined for sonicated Ethosomes. Vesicular size was found to be in the range of 0 - 5.483μm. Vesicular size was reduced up to 3 folds by sonication. In-vitro release was carried out using dialysis membrane. The values of drug release were EF₁ (20% alcohol) 76.89%, EF₂ (20% alcohol) 82.31%, EF₃ (20% alcohol) 73.62, EF₄ (30% alcohol) 86.42%, EF_5 (40% alcohol) 72.09%, EF_6 (50% alcohol) 63.21%. The order of drug release was found to be first order for all the formulations. Percentage drug accumulation into skin was also found to be maximum by the Ethosomes containing 30 % w/w ethanol and 3% lecithin which showed effective subdermal deposition and indicated better subdermal action for hypertension

Keywords: Trans dermal, lipid bilayer, alcohol, lecithin, Ethosomal system

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1. Introduction

Optimization of drug delivery through human skin is important in modern therapy. Recently, the transdermal route vied with oral treatment as the most successful innovative research area in drug delivery¹.Transdermal drug delivery system was first introduced more than 20 years ago. Transdermal drug delivery system is a type of convenient drug delivery system where drug goes to the systemic circulation through the protective barrier i.e. Skin. Only the lipophilic drugs having molecular weight < 500 Daltons can pass through it. Ethosomes have been found to be much more efficient in delivering drug to the skin².

Diacerein is a prodrug which is metabolized to rhein. It is currently approved in France for the treatment of osteoarthritis although the use of diacerein is restricted due to the side effects including severe diarrhea. Diacerein is under investigation for the treatment of Insulin Resistance, Diabetes Mellitus (Type 2), and Diabetes-Related Complications. Chemically named as 4,5-bis(acetyloxy)-9,10-dioxo-9,10-dihydroanthracene-2-carboxylic acid.

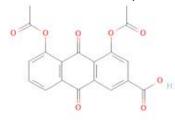
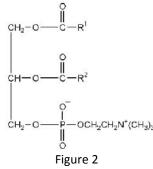


Figure 1

Lecithin 8002-43-5 1,2-diacyl-sn-glycero-3-phosphocholine (trivial chemical name, Phosphotidyl choline). The composition of lecithin (and hence also its physical properties) varies enormously depending upon the source of the lecithin and the degree of purification. Egg lecithin, for example, contains 69% Phosphotidyl choline and 24% Phosphotidyl ethanolamine, while soybean lecithin contains 21% Phosphotidyl choline, 22% Phosphotidyl ethanolamine, and 19% Phosphotidyl inositol, along with other components.



Propylene glycol: 1, 2-Dihydroxypropane; E1520; 2hydroxypropanol; methyl ethyl- ene glycol; methyl glycol; propane-1,2-diol. Propylene glycol has become widely International Journal of Medicine and Pharmaceutical Research

used as a solvent extractant, and preservative in a variety of parenteral and nonparenteral pharmaceutical formulations. It is a better general solvent than glycerin and dissolves a wide variety of materials, such as corticosteroids, phenols, sulfa drugs, barbiturates, vitamins (A and D), most alkaloids, and many local anesthetics.



Figure 3

2. Methodology Scanning of model drug (Diacerein)

10 mg of pure model drug (Diacerein) was dissolved in water and was diluted to give concentration of $10\mu g/ml$ and was scanned between 190 nm to 380 nm fort the determination of λ_{max} . The wavelength of 252 nm was selected as for λ_{max} . The same was used for further analysis of drug solution and absorbance of final standard solution was also measured at 252 nm.

Preparation of pH 6.8 phosphate buffer⁴⁶

Dissolve 28.8g of disodium hydrogen phosphate and 11.45g of potassium Di hydrogen phosphate in distilled water and then make up the volume to 1000ml with distilled water.

Preparation of calibration curve

10mg of pure Diacerein drug was taken in a 10ml standard flask and dissolved in distilled water. The volume of stock solution was made up to 10 ml with pH 6.8 phosphate buffer. From the above stock solution, 1 ml was transferred into a 10ml volumetric flask and volume was adjusted to 10 ml that corresponded to $100\mu g/ml$ Diacerein in solution. From that solution different aliquots of 0.2, 0.4, 0.6, 0.8, 1.0 and 1.2 ml were transferred to 10ml volumetric flask and volume was adjusted to 10ml with ph 6.8 phosphate buffer, which gave a concentration of 2, 4, 6, 8,10 and 12 $\mu g/ml$ respectively of final standard.

Compatibility Studies

IR spectroscopy can be used to investigate and predict any physicochemical interactions between different components in a formulation and therefore it can be applied to the selection of suitable chemically compatible excipients. The aim of present study was to test whether there is any interaction between the carriers and the drug.

Preparation of Diacerein Ethosomes

Preparation of Diacerein Ethosomes was followed by method suggested by Touitou et al., with little modification^{10.} The Ethosomal system of Diacerein comprised of

- 2-5% phospholipids (soy lecithin),
- 20-50% ethanol,
- 10% of propylene glycol,
- 0.005g of cholesterol
- Aqueous phase to 100% w/w

100mg of Diacerein was dissolved in 6ml of water in a vessel and cholesterol was added to it with vigorous

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stirring. Propylene glycol was also added during stirring. The contents were heated to 30° c. In another closed vessel, soy lecithin was dissolved in ethanol with continuous stirring and heated to 30° c. When both the solutions reached to same temperature slowly ethanol solution was added drop wise in the centre of vessel containing drug mixture. Then the stirring was continued for another 10min in a covered vessel. Water was added to adjust the volume up to 20 ml. The vesicle size of Ethosomal formulation can be decreased to desire extent using sonication³¹ or extrusion³² method. Finally the formulation was

stored under refrigeration³³. Ethosomes were formed spontaneously by this process.

Preparation of Diacerein Ethosomal gel

The best achieved Ethosomal vesicle suspension formula EF_4 was incorporated into Carbapol gel (1%, 1.5%, 2% w/w). The specified amount of Carbapol-934 powder was slowly added to pure water and kept at $100^{\circ}c$ for 20min. Triethanolamine was added to it drop wise. Appropriate amount of formula EF_4 containing Diacerein was then incorporated into gel-base. Sufficient water was finally added with other formulation ingredients with continuous stirring until homogenous formulation was achieved (G₁, G₂ and G₃). Gel containing free Diacerein drug (100mg) was also prepared by similar method using 1.5% Carbapol.

| Ethosomal formulation (EF) | Lecithin (%) | Propylene Glycol (%) | Ethanol (%) | Cholesterol (mg) | Drug (mg) | Water |
|-------------------------------|-----------------|-------------------------|----------------|---------------------|-----------|-------|
| EF ₁ | 2 | 10 | 20 | 0.05 | 100 | Q.s |
| EF ₂ | 3 | 10 | 20 | 0.05 | 100 | Q.s |
| EF ₃ | 4 | 10 | 20 | 0.05 | 100 | Q.s |
| EF ₄ | 3 | 10 | 30 | 0.05 | 100 | Q.s |
| EF ₅ | 3 | 10 | 40 | 0.05 | 100 | Q.s |
| EF ₆ | 3 | 10 | 50 | 0.05 | 100 | Q.s |
| *EF ₇ | - | 10 | 30 | 0.05 | 100 | Q.s |

Table 1: Composition of different Ethosomal formulations

*EF7- Free suspension without vehicle forming agent.

| Gel | Diacerein Ethosomal | Carbapol | Tri ethanol amine | Water |
|--------------------------------|---------------------|----------|-------------------|-------|
| formulation | suspension(ml) | (%) | (ml) | |
| EF_4G_1 | 20 | 1 | 0.5 | Q.s |
| EF ₄ G ₂ | 20 | 1.5 | 0.5 | Q.s |
| EF_4G_3 | 20 | 2 | 0.5 | Q.s |
| *G4 | 0.025g | 1.5 | 0.5 | Q.s |

Table 2 Composition of different Ethosomal gel formulation

*G-4 free drug gel

3. Results and Discussion

Calibration curve in water (make up with ph 6.8 phosphate buffer): Standard solutions of different concentrations were prepared and their absorbance was measured at 234 nm (Table 3). Calibration curve was plotted against drug concentrations versus absorbance as given in the (Figure.4).

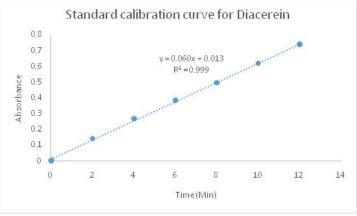


Figure 4: Standard graph of Diacerein

M. Kishore Babu et al, Int. J. Med. Pharm. Res., 2022, 10(1): 12-18 Table 3 Determination of λ_{max} of Diacereinin methanol-- λ_{max} = 252 nm

| Concentration (µg /ml) | Absorbance |
|------------------------|------------|
| 0 | 0 |
| 2 | 0.14 |
| 4 | 0.269 |
| 6 | 0.383 |
| 8 | 0.496 |
| 10 | 0.618 |
| 12 | 0.740 |

Table 4 Assessment of peaks for functional groups

| S. No. | Functional | Range of groups (drug) | Assessment peak of pure | Assessment peak of |
|--------|-------------|------------------------|---------------------------|---------------------------------|
| | groups | (cm ⁻¹) | drug (cm ⁻¹) | Ethosomal gel |
| | | | | formulation (cm ⁻¹) |
| 1 | Alkyl C-H | 2950 - 2850 (/cm) | 2697-2980cm ⁻¹ | 2507-2851 cm ⁻¹ |
| | Stretch | | | |
| 2 | Alkenyl C-H | 3010 - 3100 (/cm) | 3028-3123cm ⁻¹ | 2981 -3010 cm ⁻¹ |
| | Stretch | | | |
| 3 | Alkenyl C=C | ~3300(/cm) | | |
| | Stretch | | | |
| 4 | Amine N-H | 3500 – 3300 (/cm) | | |
| | Stretch | | | |

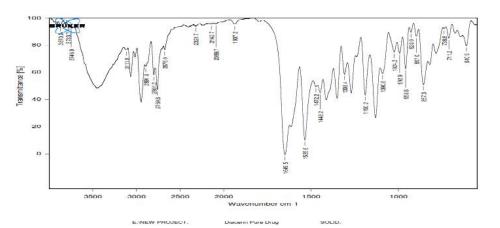
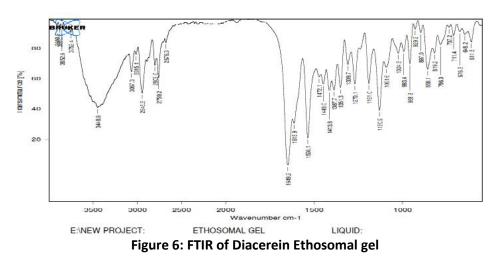


Figure 5: FTIR of pure Diacerein drug



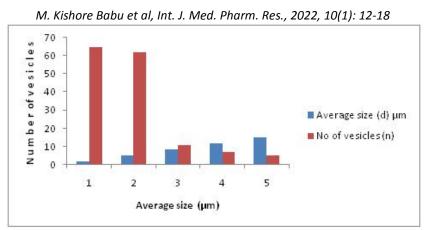


Figure 7 Size distribution range of EF₁

| Table 5 Size distribution of Ethosomal for | rmulation#2 FF | (3% Locithin | 20% ethanol) |
|---|--------------------|----------------|--------------|
| Table 5 Size distribution of Ethosoliar for | 1111UIali011#2 EF2 | (3/0 Letitiii) | |

| SIZE R | ANGE | | | | |
|-------------------------------|------------|------------------------|-----------------------|----------------------------------|-------------|
| Eye piece micrometer In μm | | Average size (d) μm | No of vesicles (n) | % No of vesicles (n/150 *100) | n x d |
| 0-1 | 0.00-3.33 | 1.665 | 60 | 40.000 | 99.9 |
| 1-2 | 3.33-6.66 | 4.995 | 45 | 30.000 | 224.775 |
| 2-3 | 6.66-9.99 | 8.325 | 30 | 20.000 | 249.75 |
| 3-4 | 9.99-13.32 | 11.655 | 10 | 6.667 | 116.55 |
| 4-5 13.32-16.65 | | 14.985 | 5 | 3.333 | 74.925 |
| | | | Σn = 150 | | Σnd = 765.9 |

Average diameter (d avg) = $\frac{\Sigma n d}{\Sigma n}$ = 5.106 µm

Table 6 Drug entrapment efficiency of Diacerein Ethosomal gel

| Formulation code | Entra | MEAN | | |
|------------------|-------|-------|-------|-------|
| EF1 | 72.19 | 71.75 | 71.82 | 71.92 |
| EF2 | 66.91 | 67.12 | 68.53 | 67.52 |
| EF3 | 60.05 | 60.00 | 60.01 | 60.02 |
| EF4 | 79.91 | 79.62 | 79.33 | 79.62 |
| EF5 | 58.01 | 55.96 | 54.96 | 56.31 |
| EF6 | 39.39 | 42.32 | 42.76 | 41.49 |

Table 7 In-vitro cumulative % drug release profile for Diacerein Ethosomes

| | Cumulative % drug release | | | | | | | | |
|-------|---------------------------|-----------------|-------|--------|-----------------|-----------------|-----------------|-----------|----------|
| Time | EF ₁ | EF ₂ | EF₃ | EF_4 | EF ₅ | EF ₆ | EF ₇ | EF_4G_2 | F_4G_4 |
| 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 30min | 14.56 | 17.5 | 12.09 | 18.09 | 11.39 | 10.01 | 11.21 | 17.82 | 8.46 |
| 2hr | 25.62 | 31.52 | 23.26 | 32.51 | 20.21 | 18.09 | 23.62 | 22.31 | 15.31 |
| 4hr | 30.1 | 38.3 | 28.21 | 39.42 | 25.22 | 22.31 | 29.72 | 28.81 | 19.71 |
| 6hr | 33.08 | 42.52 | 32.71 | 46.42 | 30.09 | 27.69 | 33.32 | 32.71 | 23.04 |
| 8hr | 38.62 | 47.81 | 37.21 | 49.31 | 36.21 | 30.03 | 37.29 | 37.61 | 25.31 |
| 10hr | 40.07 | 54.32 | 39.05 | 56.51 | 37.02 | 33.61 | 40.25 | 41.62 | 28.32 |
| 12hr | 43.62 | 58.50 | 43.02 | 60.41 | 42.3 | 37.32 | 47.91 | 46.32 | 33.13 |
| 14hr | 49.30 | 61.32 | 48.05 | 63.21 | 46.31 | 40.65 | 52.41 | 50.02 | 37.04 |
| 16hr | 52.41 | 64.92 | 51.51 | 66.72 | 49.85 | 43.32 | 52.56 | 57.42 | 39.21 |
| 18hr | 59.32 | 69.07 | 58.37 | 71.46 | 57.31 | 48.32 | 52.56 | 60.21 | 40.32 |
| 20hr | 66.02 | 75.41 | 66.42 | 76.32 | 65.21 | 52.09 | 52.56 | 66.32 | 43.01 |
| 22hr | 70.52 | 78.2 | 70.31 | 82.31 | 68.71 | 56.31 | 52.56 | 72.02 | 45.32 |
| 24hr | 76.89 | 82.31 | 73.62 | 86.42 | 72.09 | 63.21 | 52.56 | 75.62 | 47.62 |

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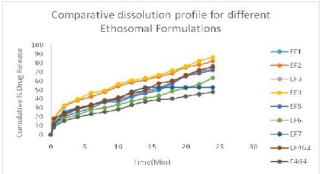


Figure 8 In-vitro drug release studies of different Ethosomal formulations

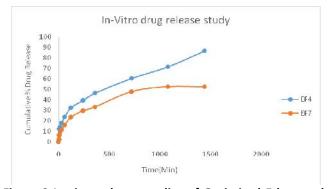


Figure 9 In-vitro release studies of Optimized Ethosomal suspension formulation (EF4) and free drug solution (EF7)

4. Conclusion

From the present study it can be concluded that Ethosomal gel is promising route of drug administration. Even though the TDDS faces the problem of drug permeation because of the rigid stratum corneum, it can be overcome by the use of penetration enhancers such as ethanol. The size of the ethosomes can be reduced by sonication thereby improving the skin permeation properties of ethosomes. By encapsulating Diacerein into ethosomes the frequency of dosing can be reduced as ethosomes cause the delivery of drug for almost 24hrs. Since the overall drug administered is reduced, the adverse drug reactions of Diacerein such as dizziness, allergy, hypotension, etc can also be reduced. Finally it can be said that many drugs such as NSAIDS, anti hypertensive drugs, anti fungal drugs, anti HIV agents, etc have the scope to be incorporated into ethosomes and thereby enhancing their activity.

5. References

- Barry BW. Novel mechanism and devices to enable successful transdermal drug delivery. European J Pharm Sci 2004; 14: 101 - 114.
- [2] Pavan Kumar. K; Radhika.P. R; Sivakumar.T. International Journal of Advances in Pharmaceutical sciences- Vol 1, No.2 (2010)
- [3] Jain N, Talegonkar S, Jain NK. New ways to enter the blood stream: Emerging strategies in

transdermal drug delivery. The Pharma Review. Sep - Oct 2004; 41-60.

- [4] Drug delivery systems by Vinayak (Vasant) V. Ranade- 2002: 199-201, 205
- [5] Modern Pharmaceutics, Fourth edition, Basic principles and systems by Gilbert S. Banker, Christopher T. Rhodes-2002: 200
- [6] Jain NK. Advances in controlled and novel drug delivery, New Delhi: CBS publication; 2001; 428 -451.
- [7] Jain S, Bhandra D, Jain S and Jain NK. Transfersomes- A novel carrier for effective transdermal drug delivery. Controlled and novel drug delivery 1 st Edition. CBS publishers and distributors New Delhi 1997: 426 - 451.
- [8] Vyas SP, Khar RK. Controlled drug delivery concepts and advances. Vallabh prakashan New Delhi. First Edition 2002: 173 - 243.
- [9] Tuitou E, Godin B, and Weiss C. Enhanced delivery into and across the skin by Ethosomal carriers. Drug Dev Research 2000; 50; 406 - 445.
- [10] Touitou E, Dayan N, Bergelson L, Godin B and Eliaz M. Ethosomes – novel vesicular carrier for enhanced delivery: characterization and skin penetration properties. J Con Release 2000; 65: 403 - 418.
- [11] Jain S, Umamaheshwari RB, Bhadra D and Jain NK. Ethosomes: A novel vesicular carrier for enhances transdermal delivery of an Anti HIV agent. Indian J Pharm Sci 2004; 66 (1): 72 - 81.
- [12] Reddy N, Pandey S and Udupa N. Improvement in bioavailability of flubiprofen in topical delivery. Ind J Hosp pharm 1993, 30: 113 - 114.
- [13] Satturwar PM, Khandare JN and Nanda VS. Niosomal delivery of Ketoconazole. Ind Drugs 2001 April: 38 (12): 620 - 624.
- [14] Amit Bhatia, Rajiv Kumar and Om Prakash Katare. Tamoxifen in topical liposomes: development, characterization and in -vitro evaluation. J Pharm Sci 2004; 7 (2): 252 - 259.
- [15] Kaplun-Frischoff Y and Touitou E. Testosterone skin permeation enhancement by method through formation of eutectic with drug and interaction with skin lipids. J Pharm Sci 1997; 86: 1394 - 9.
- [16] Touitou E, Godin B, Dayan N, Weiss C, Piliponsky A and Levi-Schaffer f. Intracellular delivery mediated by ethosomal carrier. Biomaterials 2001; 22: 3053-3059.
- [17] Chetoni P, Rossi S, Burgalassi S, Monti D Mariotti and Saettone MF. Comparison of liposomes-Encapsulated acyclovir with acyclovir ointment: ocular phamacokinetics in rabbits. J Ocular Pharmaco Thercop 2004 Apr; 22 (2): 169 - 177.
- [18] Lboutounne H, Guillaune YC, Michel L, Humbert PH and Millet J. Study and development of

encapsulated forms of 4, 5, 8 - Trimethyl prosaten for topical drug delivery. Drug Dev Res 2004, Feb; 61 (2): 36 - 94.

- [19] Lodzki M, Godin B, Rakou L, Mechoulan R, Gallily R and Touitou E. Cannabidiol - transdermal delivery and anti-inflammatory effect in a murine model. J Control Rel 2003; 93: 377 - 387.
- [20] Horwitz E, Pisanty S, Czerniski R, Helser M, Eliav E and Touitou E. A clinical evaluation of a novel liposomal carrier for acyclovir in the topical treatment of recurrent herpes labialis. Oral Medicine 1999 Feb; 87: 700 - 708.
- [21] Godin B and Touitou E. Mechanism of Bacitracin permeation enhancement through the skin and cellular membranes from and ethosomal carrier. J Control Rel 200; 94: 365 - 379.
- [22] Drug Bank- Open data drug and Drug target database
- [23] Junginger HE, Hofland HE and Bouwrtra JA. Pharm Ztg 1991; 136: 9 - 24.
- [24] Natasa Skalko, Cajokovac M. and Jalsenjak I. Liposomes with clindamycin hydrochloride in the therapy of acne vulgaris International Journal of Pharmaceutics 1992; 97 - 101.
- [25] Raja Naresh RA, Singh UV, Udupa N and Pillai GK. Encapsulation of diclofenac sodium in niosomes. Ind Drug 1993; 30; 275 – 276
- [26] Reddy N, Pandey S and Udupa N. Improvement in bioavailability of flubiprofen in topical delivery. Ind J Hosp pharm 1993, 30: 113 – 114
- [27] Bhavana Vora, Khopade AJ and Jain NK. Niosome based transdermal system for contraceptive delivery. IPC Abstract 1996: AP59; 41.
- [28] Arardottir HB, Sveinsson SJ and Kristmundsdotir T. The release of clindamycin phosphate from a suspension of different types of liposomes and selected topical dosage forms. Int J Pharm 1996; 134: 71 - 77.
- [29] Khandare JN, Jiwandad BH and Uppal RR. Preparation and evaluation of nimesulide niosomes for topical delivey. Int Drug 2001 April; 38 (4): 197 - 202.
- [30] Braun FO, Kortung HC and Maibach Griebanch conference: Liposomes Dermatics, Springarverlag. Berlin, Herdelberg 1992
- [31] Berner B and Liu P Alcohols, in Smith EW and Mailbach H I. Percutaneous penetration enhancers. CRC Press, Boca Raton 1995: 45 - 60.
- [32] Koskela RV, Kirjavainen M, Monkkonen J, Urthi A and Kiesvaara J. Enhancement of percutaneous absortion of naproxen by Phospholipids. Int J Pharm 1998; 175: 225 - 230.

- [33] Gondaliya DP and Pundarikakasudu K. Studies in formation, characterization of transdermal permeation of nemesulide from aqueous and emulgel. Ind Drug 2002: 39 (9): 465 – 473.
- [34] Sang-Chul S, Hee-Jung K, In-joon O, Cheong-weon C and Kyu-ho Y. Development of tretinoin gels for enhanced transdermal delivery. Eur J Pharma Biopharm 2005 may; 60 (1): 67 71.
- [35] Patel MM, Sheth MN and Harinawala AI. Studies in the Transdermal formulations of metaprolol tartrate and their evaluation using human cadaver skin. The East Pharmacist 1993 Dec: 129 - 131.
- [36] Indian Pharmacopoeia Addendum 2010, Volume 2 published by The Indian Pharmacopoeia Commission, Ghaziabad: 76, 1607 - 1610.
- [37] Goodman and Gilman's Manual of Pharmacology and Therapeutics by Laurence L. Brunton, Keith L.Parker: 525-527.