



International Journal of Medicine and Pharmaceutical Research

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RESEARCH ARTICLE

Evaluation of *in-vitro* Antitumor Property of Water Weeds (*Eichhorniacrassipes* and *Pistiastratiotes*) by MTT Assay

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ABSTRACT

Abstract: The water weeds *Eichhorniacrassipes* and *Pistiastratiotes* were selected to investigate the antitumor property *in-vitro* by MTT Assay. The IC₅₀ (IC-Inhibitory Concentration) was calculated for all the fractions of the selected aquatic plants isolated through column chromatography. The fractions were subjected for HPLC analysis. At higher concentration of drug (500mg) less viable cells were found. Pistia have shown better cytotoxicity than Eichhornia when subjected to B16F10 cells, the % of viable cells scored at 500mg of aqueous fractions was 26 and 35 respectively. The IC₅₀ of HPLC eluted fractions of Eichhornia & Pistia was found between 200 and 500mg/ml but the crude extracts of these water weeds did not respond similarly. The crude and alcoholic fractions of *Eichhorniacrassipes* and aqueous and methanolic fractions of *Pistiastratiotes* proved as potential tumor inhibitor.

Keywords: MTT, cytotoxicity, antitumor, *in-vitro*.

ARTICLE INFO

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PAPER QR-CODE

ARTICLE HISTORY: Received 19 June 2018, Accepted 21 July 2018, Available Online 10 August 2018

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Citation: N. Ganesh, et al. Evaluation of *in-vitro* Antitumor Property of Water Weeds (*Eichhorniacrassipes* and *Pistiastratiotes*) by MTT Assay. *Int. J. Med. Pharm. Res.*, 2018, 6(4): 138-147.

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1. Introduction

Several plants containing flavonoid derivatives have found application as disease preventive and therapeutic agents in traditional medicine in Asia for thousands of years. The International Journal of Medicine and Pharmaceutical Research

much lower risk of colon, prostate and breast cancers in Asians who consume more vegetables and fruits may indicate that flavonoid components act as natural chemo

preventive and anticancer agents. Experimental animal studies have indicated that certain dietary flavonoids possess antitumor activity (Cithan Kanadaswami *et al.*, 2005). The documented biological effects of flavonoids include anti-inflammatory, anti-allergic, antimicrobial, hepatoprotective, antiviral, antithrombotic, cardioprotective, capillary strengthening, antidiabetic, anticarcinogenic and antineoplastic effects, among others (Harborne JB & Williams CA, 2000; Middleton E Jr *et al.*, 2000; Kandaswami C & Middleton E Jr, 1994; Middleton E Jr & Kandaswami C, 1992; Middleton E Jr & Kandaswami C, 1994; Castillo MH *et al.*, 1989; Huang YT *et al.*, 1999; Kappagoda CT, 2005). Certain plant compounds, such as isoflavonoids, flavonoids and lignans, are receiving much attention as alleged cancer protective agents in populations with low incidences of breast and prostate cancer (Adlercreutz H *et al.*, 1991).

Several flavonoids not only inhibit the growth of tumor cells (Middleton E Jr *et al.*, 2000; Huang YT *et al.*, 1999; Soulinna EM *et al.*, 1975; Scambia G *et al.*, 1990; Kandaswami C *et al.*, 1991) but also induce cell differentiation (Constantinou A *et al.*, 1990). Earlier reports indicate that some flavonoids also show potent antitumor activity *in-vivo* (Edwards JM *et al.*, 1979; Molnar J *et al.*, 1981). MTT assay was performed to evaluate the antitumor properties of the water weeds. The MTT (3-[4,5-dimethylthiazol-2-yl]-2,5 diphenyltetrazolium bromide) assay is based on the conversion of MTT into formazan crystals by viable cells which determines mitochondrial activity.

Generally the total mitochondrial activity is related to the number of viable cells, therefore this assay is used to measure the *in-vitro* cytotoxic effects of drugs on cell lines or primary patient cells (Van Meerloo *et al.*, 2011). The basis of this assay is the reduction of the soluble yellow MTT tetrazolium salt to a blue insoluble formazan product by mitochondrial succinic dehydrogenase (Mosmann T, 1983). This assay has also been used to detect cytotoxic lymphokines produced by human lymphocytes (Green *et al.*, 1984) and to measure cell activation (Gerlier and Thomasset, 1986).

2. Materials and Method

In-vitro tumor cell viability (IC₅₀) was performed through MTT cytotoxicity assay. B16F10 cells were cultured *in-vitro* and subjected for monolayer growth. The tumor cell B16F10 melanoma cytotoxicity and tumor inhibitory property was scored by MTT assay. This assay was carried out as per the standard procedure described by (Fischer *et al.*, 2003; Dutta *et al.*, 2008) with slight modifications. Different concentrations from 25mg/ml–500mg/ml of *Eichhorniacrassipes* crude extract, *Eichhornia* crystals, methanolic fraction and aqueous fraction were taken. The cells were grown in fresh culture medium supplemented with 10% FBS and 1% Penicillin-Streptomycin mixture. The growth inhibition behavior of samples were evaluated by 3-(4, 5-di-methylthiazol-2-yl)-2, 5-diphenyl tetrazolium bromide (MTT) cytotoxicity assay. The cells were seeded

evenly into 96-well flat-bottomed tissue culture plate at 5×10^3 cells/well concentration and incubated for 24hrs in a humidified atmosphere of 5% CO₂ at 37°C and treated with the formulations. Samples were applied as freshly prepared solutions in dimethylsulphoxide (DMSO). After 24 hours, 20ml of 5mg/ml MTT solution in PBS (pH 7.4) was added to each well and the plate was incubated for 2 hours at 37°C allowing viable cells to reduce the MTT into purple colored formazan crystals. The absorbance was measured at 570 nm with the help of an ELISA plate reader.

3. Result and Observation

The tumor cell B16F10 melanoma cytotoxicity was performed by MTT assay. According to the Table 4 to 9, the methanolic fraction of *Eichhornia* have shown better cytotoxicity than *Pistia* when subjected to B16F10 cells. The percentage of viable cells scored at 500mg was found to be better in all the isolates where the viability of cells found to be less than 40%. Gallic acid, Catechin, Epicatechin of methanolic fraction of *Eichhorniacrassipes* have shown 28%, 33%, 35% of viable cells respectively whereas Gallic acid, Rutin of aqueous fraction of *Eichhorniacrassipes* have shown 36% and 38% viable cells respectively at 500mg/ml dose. Rutin of crude extract of *Eichhorniacrassipes* have shown 37% of viable cells whereas Rutin1 of aqueous fraction of *Eichhorniacrassipes* and Gallic acid of methanolic fraction of *Pistia* have shown 38% and 33% viable cells respectively.

Epicatechingallate, Ellagic acid, Rutin2, Quercetin, Kaempferol and Apigenin (Cunn, Kevin, 2000) have shown less than 40% viable cells in *Eichhorniacrassipes* and *Pistia*. Apigenin, unknown of crude extract of *Eichhorniacrassipes*, unknown 1, unknown 2 of aqueous extract of *Eichhorniacrassipes*, unknown and 1-O-Galloyl-B-D-Glucose of aqueous fraction of *Pistia* and unknown of methanolic fraction of *Pistia* could not able to present 50% of tumor cell death even at 500 mg/ml of dose. The IC₅₀ of HPLC eluted fractions of *Eichhornia* & *Pistia* have found between 200 and 500 mg/ml but the crude extracts of these water weeds could not able to present same response (Table 1 and 2). For *Eichhornia* crystal & methanolic fractions, the IC₅₀ was at 400 mg/ml where as the *Pistia* acetone fraction have not shown any 50% cell death even at 500 mg/ml. However the Methanolic and aqueous fractions have shown IC₅₀ at 400 mg/ml.

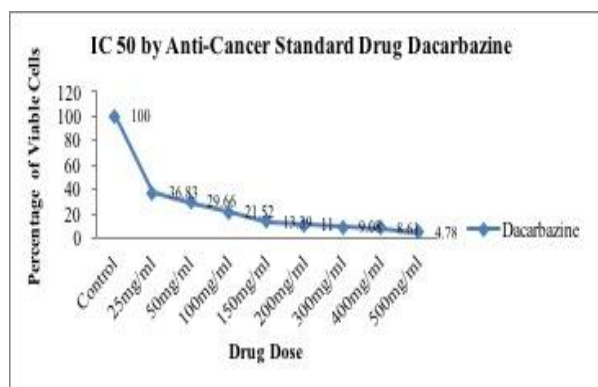


Fig.1: IC₅₀ by Anti-Cancer Standard Drug Dacarbazine

Table 1: MTT Assay of Eichhornia against B16F10 Cells

Dose	E. Crude (OD)	Viable Cells (%)	E. Crystal (OD)	Viable Cells (%)	EF1 (OD)	Viable Cells (%)	EF2 (OD)	Viable Cells (%)
Control	2.09	100	-	-	-	-	-	-
25mg/ml	1.99	95.2	2	95.69	1.91	91.38	1.8	86.12
50mg/ml	1.62	77.5	1.89	90.4	1.9	90.9	1.69	80.86
100mg/ml	1.43	68.42	1.67	79.9	1.79	88.64	1.63	77.99
150mg/ml	1.37	65.55	1.63	77.9	1.7	81.33	1.52	72.77
200mg/ml	1.07	51.96	1.44	68.89	1.5	71.77	1.41	67.46
300mg/ml	0.97	46.41	1.12	53.38	1.33	63.63	1.02	48.8
400mg/ml	0.73	34.9	0.82	38.27	0.97	46.41	0.82	39.23
500mg/ml	0.69	33.06	0.77	36.84	0.83	39.71	0.74	35.4

E. crystal- *Eichhorniacrassipes* crystals, EF1- *Eichhorniacrassipes* ethanolic fraction, EF2- *Eichhorniacrassipes* aqueous fraction, PF1- *Pistiastratiotes* acetone fraction, PF2- *Pistiastratiotes* ethanolic fraction, PF3- *Pistiastratiotes* aqueous fraction, *B16F10- (200000 cells/well)

Table 2: MTT Assay of Pistia against B16F10 Cells

Dose	Pistia Crude (OD)	Viable Cells (%)	PF1 (OD)	Viable Cells (%)	PF2 (OD)	Viable Cells (%)	PF3 (OD)	Viable Cells (%)
Control	2.09	100	-	-	-	-	-	-
25mg/ml	2.03	97.1	2.1	100	1.9	90.9	1.97	94.25
50mg/ml	1.99	95.2	1.93	92.33	1.82	87.08	1.79	85.64
100mg/ml	1.81	86.6	1.91	91.37	1.77	84.68	1.57	75.11
150mg/ml	1.82	87.08	1.82	87.06	1.66	79.42	1.32	63.15
200mg/ml	1.7	81.33	1.76	83.71	1.62	77.51	1.27	60.76
300mg/ml	1.56	74.64	1.6	76.51	1.43	68.42	1.07	51.96
400mg/ml	1.11	53.1	1.42	67.91	1.01	48.32	0.88	42.1
500mg/ml	0.82	39.22	1.37	65.88	0.83	39.71	0.56	26.79

*B16F10- (200000 cells/well)

Table 3: MTT Assay of Standard Anticancer Drug Dacarbazine against B16F10 Cells

Dacarbazine Dose	Dacarbazine (OD)	Viable Cells (%)
0.1 µl	0.77	36.83
0.2 µl	0.62	29.66
0.3 µl	0.45	21.52
0.4 µl	0.28	13.39
0.5 µl	0.23	11
0.6 µl	0.19	9.08
0.7 µl	0.18	8.61
0.8 µl	0.1	4.78

Dacarbazine Dose- 250mg/m², *B16F10- (200000 cells/well)

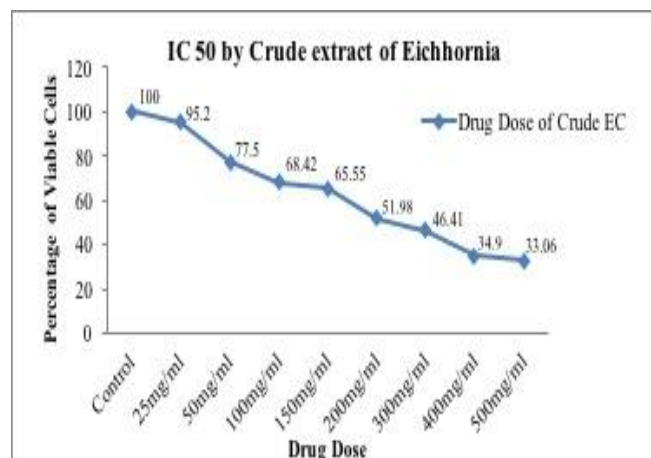


Figure2: IC 50 by Crude Extract of Eichhornia

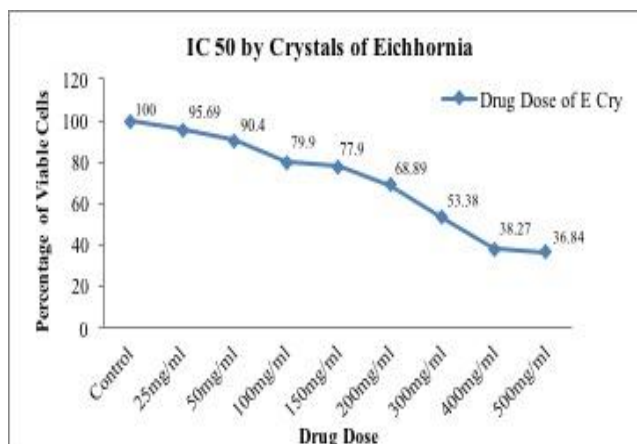


Figure3: IC 50 by Crystals of Eichhornia

Table 4: MTT Cellular Toxicity Assay of Methanolic Extract of Eichhorniacrassipes isolated through HPLC against B16F10 Cells

Sample Name	Compound Isolated	Dose Groups:		OD	Mean	% Viability
		A1, A2 – 200mg/ml	B1, B2 – 500mg/ml			
Methanolic fraction of <i>Eichhorniacrassipes</i>	Gallic acid	A1		0.704	A-0.661	A-31.6%
		A2		0.619		
		B1		0.632	B-0.599	B-28.6%
		B2		0.566		
	Catechin	A1		0.847	A-0.736	A-35.2%
		A2		0.625		
		B1		0.801	B-0.702	B-33.5%
		B2		0.603		
	Epicatechin	A1		0.604	A-0.755	A-36.1%
		A2		0.906		
		B1		0.884	B-0.745	B-35.6%
		B2		0.606		
	Epicatechingallate	A1		0.806	A-0.856	A-40.9%
		A2		0.907		
		B1		0.815	B-0.761	B-36.4%
		B2		0.708		
Apigenin	A1		0.936	A-0.857	A-41%	
	A2		0.778			
	B1		0.801	B-0.727	B-34.7%	
	B2		0.653			

*B16F10- (100000 cells/well)

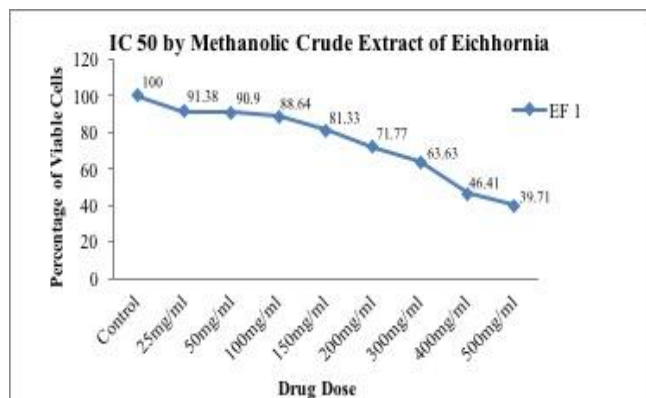


Figure 4: IC 50 by Methanolic Crude Extract of Eichhornia

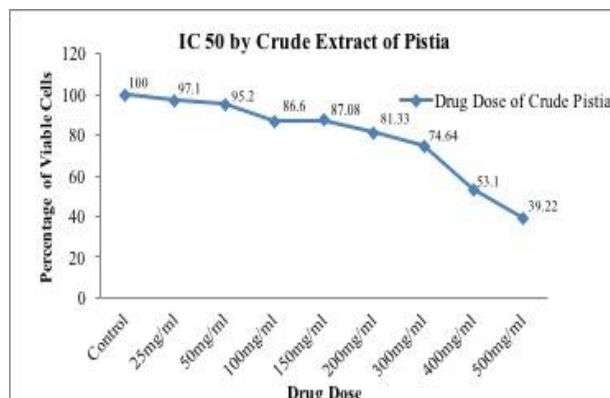


Figure 6: IC 50 by Crude Extract of Pistia

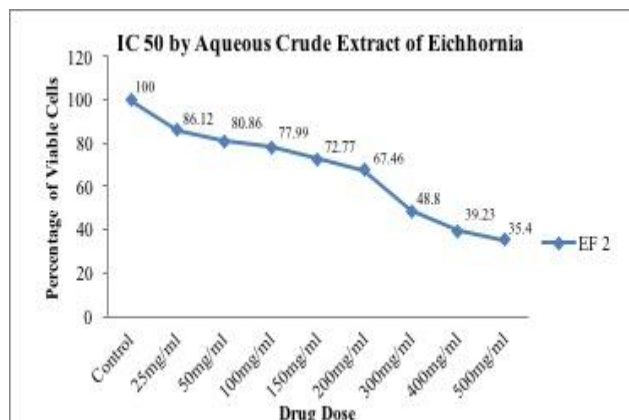


Figure 5: IC 50 by Aqueous Crude Extract of Eichhornia

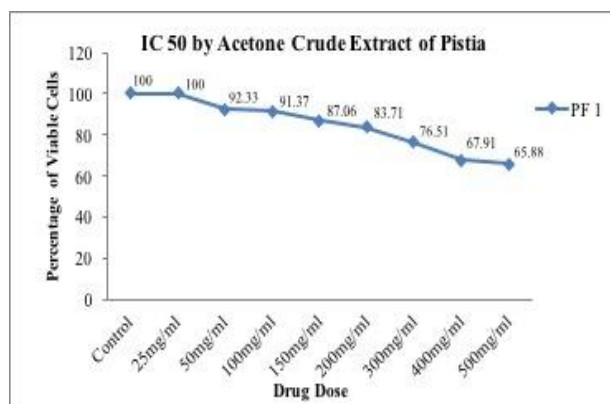


Figure 7: IC 50 by Acetone Crude Extract of Pistia

Table 5: MTT Cellular Toxicity Assay of Aqueous Extract of *Eichhorniacrassipes* isolated through HPLC against B16F10 Cells

Sample Name	Compound Isolated	Dose Groups:A1, A2 – 200mg/ml B1, B2 – 500mg/ml	OD	Mean	% Viability
Aqueous fraction of <i>Eichhorniacrassipes</i>	Gallic acid	A1	0.712	A-0.769	A-36.7%
		A2	0.826		
		B1	0.808	B-0.761	B-36.4%
		B2	0.714		
	Epigallocatechin	A1	0.778	A-0.857	A-41.0%
		A2	0.936		
		B1	0.866	B-0.849	B-40.6%
		B2	0.832		
	Protocatechuic Acid	A1	0.991	A-0.899	A-43.0%
		A2	0.808		
		B1	0.964	B-0.848	B-40.5%
		B2	0.732		
	Unknown 1	A1	1.305	A-1.134	A-54.2%
		A2	0.963		
		B1	1.234	B-1.109	B-53.0%
		B2	0.984		
	Unknown 2	A1	1.378	A-1.33	A-63.6%
		A2	1.282		
		B1	0.903	B-1.093	B-52.2%
		B2	1.283		
	Rutin 1	A1	0.809	A-0.813	A-38.8%
		A2	0.817		
		B1	0.874	B-0.801	B-38.3%
		B2	0.729		
	Rutin 2	A1	0.805	A-0.862	A-41.2%
		A2	0.92		
		B1	0.804	B-0.835	B-39.9%
		B2	0.867		
	Ellagic Acid	A1	0.929	A-0.904	A-43.2%
		A2	0.879		
		B1	0.861	B-0.92	B-44.0%
		B2	0.979		
Vallinic Acid	A1	1.145	A-1.042	A-49.8%	
	A2	0.94			
	B1	0.907	B-1.006	B-48.1%	
	B2	1.105			
Kaempferol	A1	0.984	A-0.942	A-45.0%	
	A2	0.901			
	B1	0.892	B-0.854	B-40.8%	
	B2	0.817			
Myricetin	A1	1.01	A-0.96	A-45.9%	
	A2	0.91			
	B1	0.949	B-0.900	B-43.0%	
	B2	0.852			
Quercetin	A1	0.769	A-0.866	A-41.4%	
	A2	0.963			
	B1	0.803	B-1.003	B-47.9%	
	B2	1.204			
Naringinin	A1	0.905	A-1.006	A-48.1%	
	A2	1.107			
	B1	0.869	B-0.881	B-42.1%	
	B2	0.894			

Table No. 6: MTT Cellular Toxicity Assay of Crude Extract of *Eichhorniacrassipes* isolated through HPLC Analysis against B16F10 Cells

Sample No.	Compound Isolated	Dose Groups	OD	Mean	% Viability
Crude Extract of <i>Eichhorniacrassipes</i>	Rutin	A1-200mg/ml	0.742	A-0.785 B-0.782	A-37.5% B-37.4%
		A2-200mg/ml	0.829		
		B1-500mg/ml	0.879		
		B2-500mg/ml	0.685		
	Ellagic Acid	A1-200mg/ml	0.787	A-0.848 B-0.835	A-40.5% B- 39.9%
		A2-200mg/ml	0.910		
		B1-500mg/ml	0.836		
		B2-500mg/ml	0.834		
	Vallinic Acid	A1-200mg/ml	0.912	A-1.005 B-0.985	A-48.0% B-47.1%
		A2-200mg/ml	1.098		
		B1-500mg/ml	1.039		
		B2-500mg/ml	0.932		
Kaempferol	A1-200mg/ml	0.945	A-0.887 B-0.872	A-42.4% B-41.7%	
	A2-200mg/ml	0.830			
	B1-500mg/ml	0.852			
	B2-500mg/ml	0.893			
Unknown	A1-200mg/ml	1.989	A-1.558 B-1.089	A-61.2% B-52.1%	
	A2-200mg/ml	1.127			
	B1-500mg/ml	0.901			
	B2-500mg/ml	1.278			
Myricetin	A1-200mg/ml	0.965	A-0.957 B-0.954	A-45.7% B-45.6%	
	A2-200mg/ml	0.950			
	B1-500mg/ml	0.829			
	B2-500mg/ml	1.079			
Quercetin	A1-200mg/ml	0.826	A-0.843 B-0.838	A-40.3% B-40.0%	
	A2-200mg/ml	0.860			
	B1-500mg/ml	0.849			
	B2-500mg/ml	0.828			
Astragalgin	A1-200mg/ml	0.985	A-0.891 B-0.873	A-42.6% B-41.7%	
	A2-200mg/ml	0.798			
	B1-500mg/ml	0.786			
	B2-500mg/ml	0.961			
Caffeic Acid	A1-200mg/ml	0.908	A-1.013 B-1.043	A-48.4% B-49.9%	
	A2-200mg/ml	1.118			
	B1-500mg/ml	1.109			
	B2-500mg/ml	0.978			
Apigenin	A1-200mg/ml	0.945	A-52.2% B-1.048	A-52.2% B-50.1%	
	A2-200mg/ml	1.240			
	B1-500mg/ml	1.087			
	B2-500mg/ml	1.010			
Naringinin	A1-200mg/ml	0.938	A-0.983 B-0.989	A-47.0% B-47.3%	
	A2-200mg/ml	1.028			
	B1-500mg/ml	0.997			
	B2-500mg/ml	0.982			

*B16F10- (100000 cells/well)

Table No.7: MTT Cellular Toxicity Assay of Acetone Extract of *Pistiastratiotes* isolated through HPLC against B16F10 Cells

Sample No.	Compound Isolated	Dose Groups	OD	Mean	% Viability
Acetone fraction of <i>Pistiastratiotes</i>	Gallic acid	A1-200mg/ml	0.837	A-0.864	A-41.3%
		A2-200mg/ml	0.892		
		B1-500mg/ml	0.874	B-0.801	
		B2-500mg/ml	0.729		
	Rutin	A1-200mg/ml	0.961	A-0.97	A-46.4%
		A2-200mg/ml	0.979		
		B1-500mg/ml	0.841	B-0.885	
		B2-500mg/ml	0.929		
	Vallinic Acid	A1-200mg/ml	1.034	A-1.01	A-48.3%
		A2-200mg/ml	0.986		
		B1-500mg/ml	1.019	B-0.984	
		B2-500mg/ml	0.95		

***B16F10- (100000 cells/well)**

Table No. 8: MTT Cellular Toxicity Assay of Aqueous Extract of *Pistiastratiotes* isolated through HPLC against B16F10 Cells

Sample No.	Compound Isolated	Dose Groups	OD	Mean	% Viability
Aqueous fraction of <i>Pistiastratiotes</i>	Gallic acid	A1-200mg/ml	0.817	A-0.839	A-40.1%
		A2-200mg/ml	0.862		
		B1-500mg/ml	0.837	B-0.838	
		B2-500mg/ml	0.839		
	1-O-Galloyl – -D-Glucose	A1-200mg/ml	1.55	A-1.233	A-58.9%
		A2-200mg/ml	0.916		
		B1-500mg/ml	0.982	B-1.022	
		B2-500mg/ml	1.603		
	Rutin	A1-200mg/ml	0.893	A-0.869	A-41.5%
		A2-200mg/ml	0.845		
		B1-500mg/ml	0.902	B-0.863	
		B2-500mg/ml	0.825		
	Catechin	A1-200mg/ml	1.082	A-0.945	A-45.2%
		A2-200mg/ml	0.808		
		B1-500mg/ml	0.983	B-0.906	
		B2-500mg/ml	0.829		
	Epicatechin	A1-200mg/ml	0.92	A-0.959	A-45.8%
		A2-200mg/ml	0.999		
		B1-500mg/ml	0.94	B-0.956	
		B2-500mg/ml	0.972		
	Epicatechingallate	A1-200mg/ml	0.934	A-0.990	A-47.3%
		A2-200mg/ml	1.047		
		B1-500mg/ml	0.854	B-0.981	
		B2-500mg/ml	1.109		
	Unknown	A1-200mg/ml	1.923	A-1.454	A-69.5%
		A2-200mg/ml	0.986		
		B1-500mg/ml	1.625	B-1.15	
		B2-500mg/ml	0.675		

***B16F10- (100000 cells/well)**

Table No. 9: MTT Cellular Toxicity Assay of Methanolic Extract of *Pistiastratiotes* isolated through HPLC against B16F10 Cells

Sample No.	Compound Isolated	Dose Groups	OD	Mean	% Viability
Methanolic fraction of <i>Pistiastratiotes</i>	Gallic acid	A1-200mg/ml	0.894	A-0.849	A-40.6%
		A2-200mg/ml	0.805		
		B1-500mg/ml	0.194	B-0.703	B-33.6%
		B2-500mg/ml	1.212		
	Rutin	A1-200mg/ml	0.932	A-0.894	A-42.7%
		A2-200mg/ml	0.856		
		B1-500mg/ml	0.804	B-0.890	B-42.5%
		B2-500mg/ml	0.977		
	Catechin	A1-200mg/ml	0.966	A-0.933	A-44.6%
		A2-200mg/ml	0.9		
		B1-500mg/ml	0.983	B-0.906	B-43.3%
		B2-500mg/ml	0.829		
	Unknown	A1-200mg/ml	1.076	A-1.091	A-52.2%
		A2-200mg/ml	1.106		
		B1-500mg/ml	1.101	B-1.038	B-49.6%
		B2-500mg/ml	0.976		
Kaempferol	A1-200mg/ml	0.92	A-0.959	A-45.8%	
	A2-200mg/ml	0.999			
	B1-500mg/ml	0.914	B-0.935	B-44.7%	
	B2-500mg/ml	0.956			

*B16F10- (100000 cells/well)

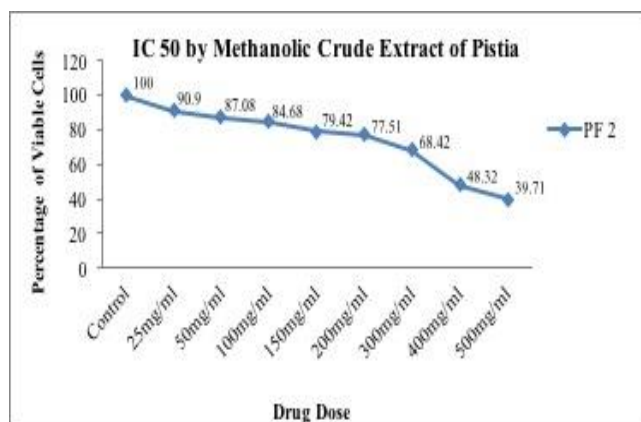


Figure 8: IC 50 by Methanolic Crude Extract of Pistia

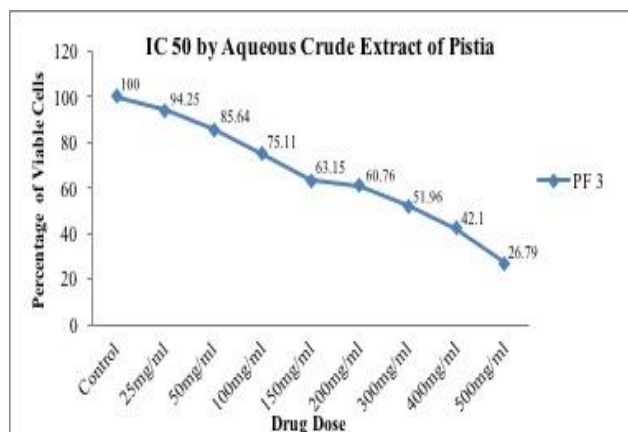


Figure 9: IC 50 by Aqueous Crude Extract of Pistia

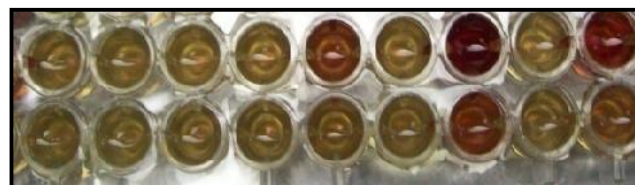
The Optical Density was screened by ELISA Plate Reader ELX-808 iu/AVL Biomedical.



Control without any drug



Test Group



Dacarbazine

Figure 10: MTT Assay for Crude Extract of *Eichhornia* and *Pistia* compared to Dacarbazine

4. Discussion and Conclusion

It was found that Eichhornia crude inhibited 46.4% of the cells and aqueous fractions inhibited 48.8% of the cell. But Eichhornia crystals and methanolic fraction inhibited 38.27% and 46.41% of tumor cells. On the other hand, Pistiamethanolic fraction and aqueous fraction were able to inhibit 50% of cancer cells at 400mg/ml dose whereas Pistia crude able to inhibit at 500mg/ml. Acetone fraction has not shown 50% cell death even 500mg/ml. The IC 50 of *Eichhorniacrassipes* and Pistia against B16F10 cells was compared with dacarbazine. At very low concentration 25mg/ml of dacarbazine was able to inhibit 36.83% of the total cells. Gallic acid, Catechin, Epicatechin, Epicatechingallate of methanolic fraction of Eichhornia have shown best tumor cell inhibiting property than methanolic fraction of Pistia. Gallic acid and Rutin 1 of aqueous fraction of Eichhornia have shown better result than aqueous fraction of Pistia.

5. Acknowledgement

The authors are thankful to Madhya Pradesh Biotechnology Council for sponsoring the project. We also acknowledge Mrs. Asha Joshi (Chairman), Dr. K.V. Pandya(Director), Dr. Pradeep Kolekar (Medical Director), Mr. Rakesh Joshi (Additional Director) and Mrs. DivyaParashar (CEO) of Jawaharlal Nehru Cancer Hospital & Research Centre, Bhopal for providing the necessary laboratory facilities and support.

Conflict of Interest: The authors declare that there are no conflicts of interest.

6. References

- [1] K Cithan, T Lung, L Ping, H Juan, Hwang, K Ferng, H Ying and L Ming. The antitumor activities of flavonoids. *In-vivo*, 2005, 19: 895-910.
- [2] JB Harborne and CA Williams. Advances in flavonoid research since 1992. *Phytochemistry*, 2000, 55: 481-504.
- [3] E Middleton, C Kandaswami and TC Theoharidis. The impact of plant flavonoids on mammalian biology: implications for inflammations, heart disease and cancer. *Pharmacol Rev*, 2000, 52: 673-751.
- [4] C Kandaswami and E Middleton. Free radical scavenging and antioxidant activity of plant flavonoids. *AdvExperimen Med Biol*, 1994, 366: 351-376.
- [5] C Kandaswami and E Middleton. Effects of flavonoids on immune and inflammatory cell function. *Biochem Pharmacol*, 1992, 43, 1167-1179.
- [6] E Middleton and C Kandaswami. Potential health-promoting properties of citrus flavonoids. *Food Tech*, 1994, 48: 115-119.
- [7] MH Castillo, E Perkins, JH Campbell, R Doerr, JM Hasset, C Kandaswami and E Middleton. The effects of the bioflavonoids quercetin on squamous cell carcinoma of head and neck origin. *Am J Surgery*, 1989, 158: 351-355.

- [8] YT Huang, JJ Hwang, PP Lee, FC Ke, JH Huang, CJ Huang, C Kandaswami, E Middleton and MT Lee. Effects of luteolin and quercetin, inhibitors of tyrosine kinase, on growth and metastasis-associated properties in A431 cells overexpressing epidermal growth factor receptor. *Br J Pharmacol*, 1999, 128:999-1010.
- [9] CT Kappagoda, M Karim, K McCormic and C Kandaswami. Unraveling the French paradox. *ChemInnov*, 2000, 30(9): 26-31.
- [10] H Adlercreutz, H Honjo, A Higashi, T Fotsis, E Hamalainen, T Hasegawa and H Okada. Urinary excretion of lignans and isoflavonoid phytoestrogens in Japanese men and women consuming a traditional Japanese diet. *Am J ClinNutr*, 1991, 54: 1093-1100.
- [11] EM Soulinna, RN Buchsbaum and E Racker. The effect of flavonoids on aerobic glycolysis and growth of tumor cells. *Cancer Res*, 1975, 35: 1865-1872.
- [12] G Scambia, FO Ranelletti, P Benedetti, M Piantelli, G Bonanno, R Vincenzo, G Ferrandina, C Rumi, LM Larocca and S Mancuso. Inhibitory effect of quercetin on OVCA 433 cells and presence of type II oestrogen binding sites in primary ovarian tumours and cultured cells. *Br J Cancer*, 1990, 62: 942-946.
- [13] C Kandaswami, E Perkins, DS Soloniuk, G Drzewiecki and E Middleton. Antiproliferative effects of citrus flavonoids on a human squamous cell carcinoma *in vitro*. *Cancer Lett*, 1991, 56: 147-152.
- [14] A Constantinou, K Kiguchi and E Huberman. Induction of differentiation and DNA strand breakage in human HL-60 and K-562 leukemia cells by genistein. *Cancer Res*, 1990, 50: 2618-2624.
- [15] JM Edwards, RF Raffauf and WL Quesne. Antineoplastic activity and cytotoxicity of flavones, isoflavones and flavanones, *J Nat Prod*, 1979, 42: 85-91.
- [16] J Molnar, I Beladi, K Domonkos, S Foldeak, K Boda and A Veckenstedt. Antitumor activity of flavonoids on NK/Lyascites tumor cells. *Neoplasma*, 1981, 28: 11-18.
- [17] J Meerloo, GJ Kaspers, J Cloos. Cell sensitivity assays: the MTT assay. *Methods MolBiol*, 2011, 731: 237-45.
- [18] T Mosmann. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J. Immunol. Meth*, 1983, 65: 55-63.
- [19] LM Green, JL Reade, and CF Ware. Rapid colorimetric assay for cell viability: application of the quantitation of cytotoxic and growth inhibition lymphokines. *J. Immunol. Meth*, 1984, 70: 257-268.
- [20] D Gerlier and N Thomasset. Use of MTT colorimetric assay to measure cell activation. *J. Immunol. Meth*, 1986, 94: 57-63.

- [21] T Dutta, M Garg, V Dubey, D Mishra, K Singh, D Pandita. Toxicological Investigation of surface engineered fifth generation poly (propyleneimine) dendrimers *in-vivo*. *Nanotoxicology*, 2008a, 2: 62–70.
- [22] D Fischer, Y Li, B Ahlemeyer, J Kriegelstein, T Kissel. *In-vitro* cytotoxicity testing of polycations: Influence of polymer structure on cell viability and hemolysis. *Biomaterials*, 2003, 24:1121.
- [23] Cunn. S Kevin, Gould, R Kenneth. Functional role of anthocyanins in the leaves of *Quintiniaserrata*. *Journal of Experimental Botany*, 2000, Vol. 51, No. 347, pp. 1107-1115.