



International Journal of Medicine and Pharmaceutical Research

Journal Home Page: www.pharmaresearchlibrary.com/ijmpr



Research Article

Open Access

A Novel Method for the Simultaneous Estimation of Imipenem and Cilastatin by Using RP-HPLC in its Bulk and Pharmaceutical Dosage Form

Satheesh thotla*, R. Vasanthi, Dr. Alagar Raja.M, K.N.V Rao, Devid Banji

Department of Pharmaceutical Analysis, Nalanda College of Pharmacy, Nalgonda, Telangana, India

ABSTRACT

A new method was established for simultaneous estimation of Imipenem and cilastatin by RP-HPLC method. The chromatographic conditions were successfully developed for the separation of Imipenem and cilastatin by using Symmetry C18 column (4.6×150mm)5 μ , flow rate was 1ml/min, mobile phase ratio was (70:30 v/v) Acetonitrile : phosphate buffer (KH₂PO₄ and K₂HPO₄) pH 3 (pH was adjusted with ortho-phosphoric acid), detection wavelength was 240nm. The instrument used was WATERS HPLC Auto Sampler, Separation module 2695, photo diode array detector 996, Empower-software version-2. The retention times were found to be 2.449 mins and 3.191 mins. The % purity of Imipenem and cilastatin was found to be 100.27% and 99.87% respectively. The system suitability parameters for Imipenem and cilastatin such as theoretical plates and tailing factor were found to be 2438, 1.26 and 2235 and 1.33, the resolution was found to be 8.67. The linearity study of Imipenem and cilastatin was found in concentration range of 50 μ g-250 μ g and 5 μ g-50 μ g and correlation coefficient (r^2) was found to be 0.999 and 0.999, % recovery was found to be 99.56% and 99.48%.

Keywords: Imepenam, Cilastatin, HPLC, Validation studies

ARTICLE INFO

CONTENTS

- | | |
|-------------------------------------|------|
| 1. Introduction | 1134 |
| 2. Materials and Methods | 1134 |
| 3. Results and discussion | 1135 |
| 4. Conclusion | 1137 |
| 5. References | 1137 |

Article History: Received 29 May 2015, Accepted 09 July 2015, Available Online 10 August 2015

*Corresponding Author

Satheesh Thotla
Department of Pharmaceutical
Analysis, Nalanda College of Pharmacy
Nalgonda, Telangana, India
Manuscript ID: IJMPR2657



PAPER-QR CODE

Citation: Satheesh Thotla, et al. A Novel Method for the Simultaneous Estimation of Imipenem and Cilastatin by Using RP-HPLC in its Bulk and Pharmaceutical Dosage Form. *Int. J. Med. Pharm, Res.*, 2015, 3(4): 1133-1138.

Copyright © 2015 Satheesh Thotla, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited.

1. Introduction

Imipenem acts as an antimicrobial through the inhibition of cell wall synthesis of various gram-positive and gram-negative bacteria. This inhibition of cell wall synthesis in gram-negative bacteria is attained by binding to penicillin binding proteins (PBPs). In *E. coli* and selected strains of *P. aeruginosa*, imipenem has shown to have the highest affinity to PBP-2, PBP-1a, and PBP-1b. This preferential binding to PBP-2 and PBP-1b results in the direct conversion of the individual cell to a spheroblast, which leads to rapid cell lysis and death without filament formation. Chemically known as (5R,6S)-3-({2-[(E)-(amino methylidene)amino]ethyl} sulfanyl)-6-[(1R)-1-hydroxyethyl]-7-oxo-1-azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid.

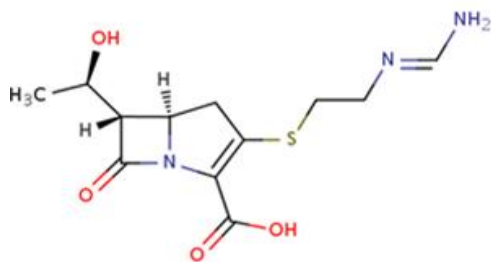


Figure 1: Imipenem

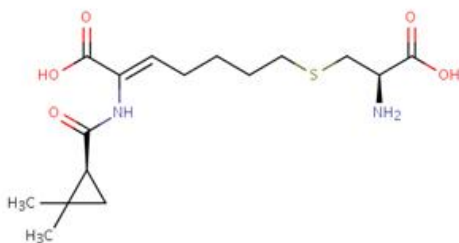


Figure 2: Cilastatin

Its Anti bacterial agent for the treatment of bacterial infections caused by susceptible and then second drug Cilastatin is a specific and reversible renal dehydropeptidase-I inhibitor. Since the antibiotic, imipenem, is hydrolyzed by dehydropeptidase-I, which resides in the brush border of the renal tubule, cilastatin is administered with imipenem to block the metabolism and thus the inactivation of imipenem so that antibacterial levels of imipenem can be attained in the urine. The drug also prevents the metabolism of leukotriene D4 to leukotriene E4 through the inhibition of leukotriene D4 dipeptidase. Chemically Known as (2Z)-7-[[{(2R)-2-amino-2-carboxyethyl} sulfanyl]-2-[(1S)-2, 2-dimethylcyclopropyl]

formamido} hept-enoinacid. Cilastatin is a competitive, reversible and specific inhibitor of dehydropeptidase-I enzyme.

2. Materials and Methods

Buffer preparation:

Preparation of Phosphate buffer :(pH: 3.0):

2.95 grams of KH_2PO_4 and 5.45 grams of K_2HPO_4 was weighed and taken into a 1000ml beaker, dissolved and diluted to 1000ml with HPLC water and pH was adjusted to 3 with ortho phosphoric acid. The resulting solution was sonicated and filtered.

Standard Preparation:

10 mg Imipenem and 10 mg Cilastatin working standard was accurately weighed and transferred into a 10ml clean dry volumetric flask and add about 2ml of diluent and sonicate to dissolve it completely and make volume up to the mark with the same solvent (Stock solution). Further pipette out 1 ml of the above stock solution into a 10ml volumetric flask and was diluted up to the mark with diluent.

Method development & Optimization:

Using Mobile phase consisting of different buffers and methanol at different concentrations and different mobile phase's pH values are attempted. The peak was observed that the shape and retention time of Imipenem and Cilastatin were found to be broad compared to the mixed phosphate buffer and acetonitrile composition of mobile phase. After selecting the best conditions based on peak performance, ACN: Phosphate buffer pH 3.0 (70: 30 % v/v) and HPLC using column is : zodiac sil RP C18 4.6×250mm 3.0 μm , the run times of the proposed method was 10 mins with isocratic solution. Column temperature is 25°C, flow rate is 1ml/min, PDA Detector is mainly used this purpose, after inject the standard solution volume was found to be 10 μL . Retention times found were about 2.365 & 3.907 mins respectively.

Method validation:

Imipenem and Cilastatin standards taken to the 25mg was accurately weighed and transferred into a 25ml of volumetric flask containing HPLC grade Methanol's diluents. It was sonicated, dissolves completely and made volume up to the mark with the same solvent. The method was validated in accordance with ICH guidelines. The parameters assessed were precision, accuracy, linearity, specificity, robustness.

Table 1: Details of Instrument

S.No	Instrument	Model No.	Software	Manufacturer's name
1	HPLC Alliance	Waters 2695	Empower	Waters
2	PDA Detector	Waters 996	UV Win 5	Lab India
3	UV double beam spectrophotometer	UV 3000	-	Satorius
4	Digital weighing balance	BSA224SCW	-	Lab India
5	pH meter	AD102U	-	-
6	Ultra sonicator	SE60US	-	-

3. Results and Discussion

A new method was established for simultaneous estimation of Imipenem and cilastatin by RP-HPLC method. The chromatographic conditions were successfully developed for the separation of Imipenem and cilastatin by using SYMMETRY C18 column (4.6×150mm) 5μ, flow rate was 1ml/min, mobile phase ratio was (70:30 v/v) Acetonitrile: phosphate buffer (KH₂PO₄ and K₂HPO₄) pH 3 (pH was adjusted with ortho phosphoric acid), detection wavelength was 240nm.

The instrument used was WATERS HPLC Auto Sampler, Separation module 2695, photo diode array detector 996, Empower-software version-2. The retention times were found to be 2.449 mins and 3.191 mins. The % purity of Imipenem and cilastatin was found to be 100.27% and 99.87% respectively. The system suitability parameters for Imipenem and cilastatin such as theoretical plates and tailing factor were found to be 2438, 1.26 and 2235 and 1.33, the resolution was found to be 8.67. The analytical method was validated according to ICH guidelines (ICH, Q2 (R1)). The linearity study of Imipenem and cilastatin was found in concentration range of 50μg-250μg and 5μg-50μg and correlation coefficient (r²) was found to be 0.999 and 0.999, % recovery was found to be 99.56% and 99.48%, %RSD for repeatability was 1.2 and 2.0, % RSD for intermediate precision was 1.1 and 1.1 respectively. LOD value was 2.17 and 6.60, and LOQ value was 0.032 and 0.1125 respectively. Hence the suggested RP-HPLC method can be used for routine analysis of Imipenem and cilastatin in API and Pharmaceutical dosage form.

Table 1

Peak Name	RT	Area	Height	USP Plate Count	USP Resolution	USP Tailing
1 Cilastatin	2.449	1214356	143778	1938.96		1.32
2 Cilastatin	2.467	1217143	139906	1847.94		1.36
3 Cilastatin	2.511	1230473	134589	1741.58		1.43
4 Imipenem	3.191	861161	88187	2438.26	3.03	1.26
5 Imipenem	3.268	855169	83700	2358.16	3.13	1.30
6 Imipenem	3.336	868604	81477	2243.15	3.05	1.36

System suitability parameters

Method Validation:

Precision:

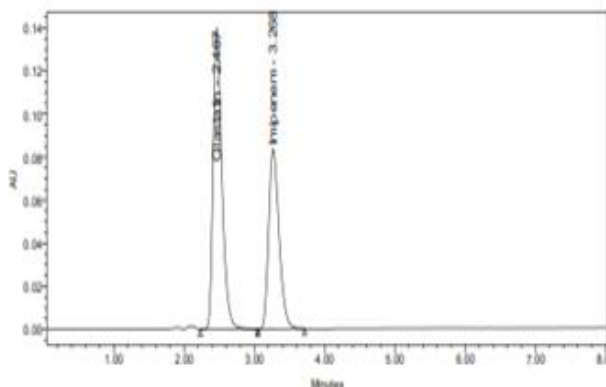


Figure 5

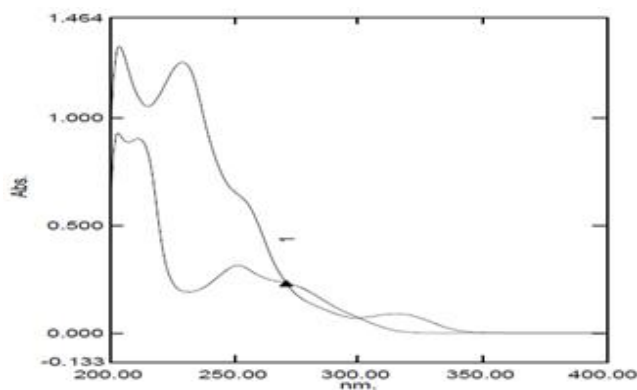


Figure 3: UV Spectrum (Isobestic Point)

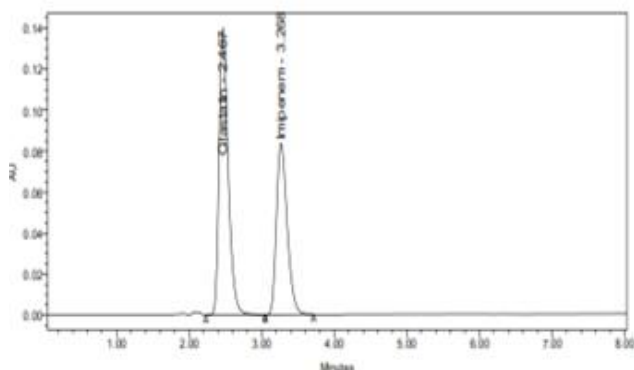


Figure 4

Table 2: Showing % RSD results for Cilastatin and Imipenem

Peak Name: Cilastatin							
	Peak Name	RT	Area	Height	USP Plate Count	USP Tailing	Injection
1	Cilastatin	3.174	1592260	146364	2973.61	1.49	1
2	Cilastatin	3.174	1600516	147364	2973.61	1.49	2
3	Cilastatin	3.323	1620416	144812	2873.05	1.52	3
4	Cilastatin	3.360	1605813	145422	2912.90	1.52	4
5	Cilastatin	3.740	1670034	136518	2654.63	1.50	5
6	Cilastatin	3.842	1571138	141267	2898.11	1.43	6
	Mean		1610030				
	Std. Dev.		33600.6				
	% RSD		2.0				

Peak Name: Imipenem								
	Peak Name	RT	Area	Height	USP Plate Count	USP Resolution	USP Tailing	Injection
1	Imipenem	4.002	1197254	141033	2852.74	4.51	1.37	5
2	Imipenem	4.673	1171214	84462	2672.17	4.42	1.38	1
3	Imipenem	4.863	1207502	84306	2557.54	4.39	1.40	3
4	Imipenem	4.920	1212400	84282	2627.38	4.24	1.38	2
5	Imipenem	5.450	1203215	82319	2444.22	4.36	1.38	6
6	Imipenem	5.637	1184663	85374	2506.06		1.32	4
	Mean		1196041.2					
	Std. Dev.		15477.3					
	% RSD		1.2					

Linearity

Table 3: Linearity Results for Imipenem

S.No	Linearity Level	Concentration	Area
1	I	50 ppm	471543
2	II	100 ppm	656277
3	III	150 ppm	794999
4	IV	200 ppm	946124
Correlation Coefficient			0.999

Acceptance Criteria: Correlation coefficient should be not less than 0.999

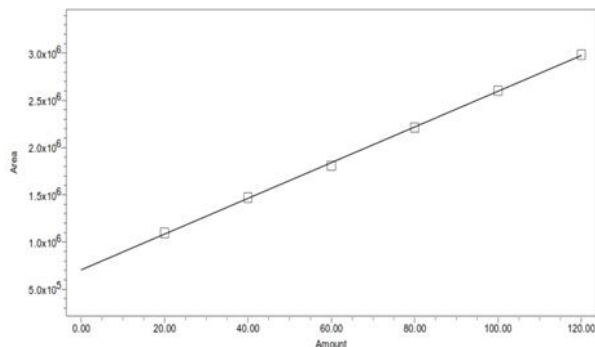


Figure 6: Showing calibration graph for Imipenem

Table 4: Linearity Results for Cilastatin

S.No	Linearity Level	Concentration	Area
1	I	5ppm	56472
2	II	10 ppm	73841
3	III	15ppm	92655
Correlation Coefficient			0.999

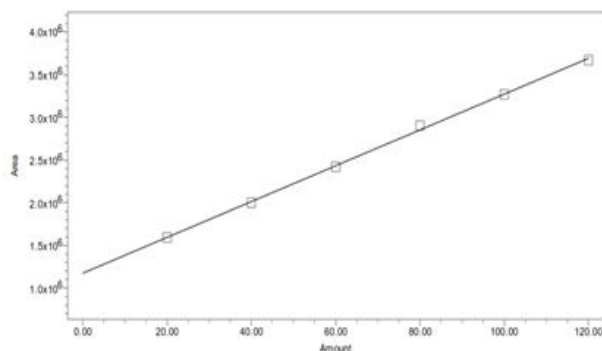


Figure 7: Showing calibration graph for cilastatin

Overlay Report

The linearity study was performed for concentration range of 50ppm-250ppm and 5ppm-50ppm of Imipenem and cilastatin and the correlation coefficient was found to be 0.999 and 0.999.(NLT 0.999).

Robustness:

The robustness was performed for the flow rate variations from 0.4ml/min to 0.6ml/min and mobile phase ratio variation from more organic phase to less organic phase ratio for Imipenem and cilastatin. The method is robust only in less flow condition and the method is robust even by change in the Mobile phase ±5% . .

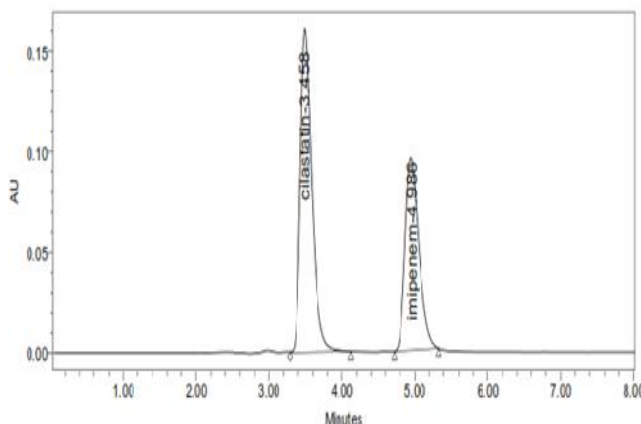


Figure 8: Chromatogram showing less flow rate 0.8ml/min

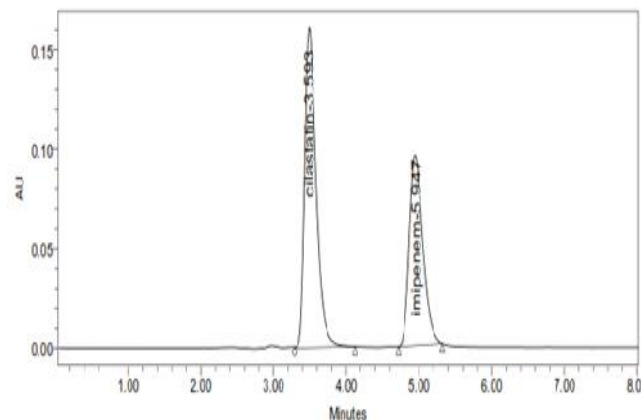


Figure 9: Chromatogram showing more flow rate 1.2 ml/min

Table 5: Showing system suitability results for Imipenem

S. No	Flow rate (ml/min)	System suitability results	
		USP Plate Count	USP Tailing
1	0.8	2590	1.39
2	1	2294	1.27
3	1.2	2146	1.26

Table 6: Showing system suitability results for Cilastatin

S. No	Flow rate (ml/min)	System suitability results	
		USP Plate Count	USP Tailing
1	0.8	5435	1.04
2	1	4891	1.03
3	1.2	4781	1.04

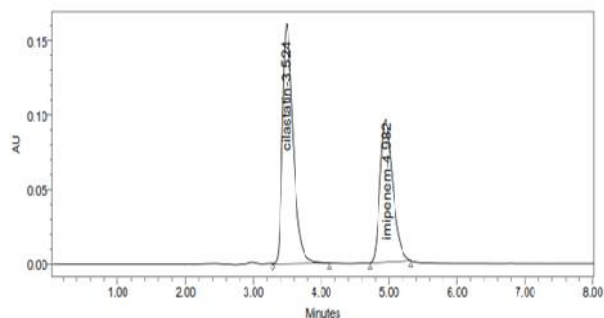


Figure 10: Chromatogram showing more organic phase ratio

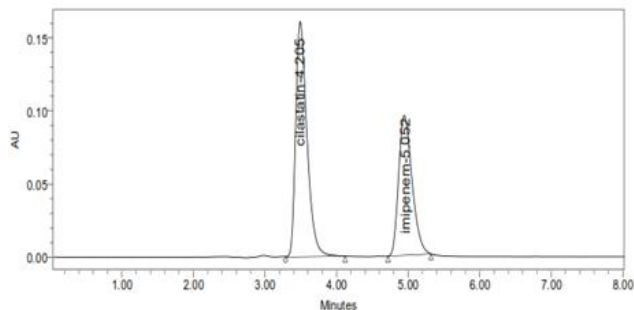


Figure 11: Chromatogram showing less organic phase ratio

Table 7: Showing system suitability results for Imipenem

S.No	Change in organic composition in the mobile phase	System suitability results	
		USP Plate Count	USP Tailing
1	5 % less	2347	1.44
2	*Actual	2294	1.27
3	5 % more	2239	1.13

Table 8: Showing system suitability results for Cilastatin

S.No	Change in organic composition in the mobile phase	System suitability results	
		USP Plate Count	USP Tailing
1	5 % less	5437	0.99
2	*Actual	4891	1.03
3	5 % more	4817	1.05

4. Conclusion

Hence it can be concluded that the proposed HPLC method is sensitive and reproducible for the determination of related substances in Imepenam and Cilastatin. The major advantage of this method was short retention time.

Table 9: Showing accuracy results for Imipenem

%Concentration (at specification level)	Average area	Amount Added (mg)	Amount found (mg)	% Recovery	Mean recovery
50%	1193706	5	4.96	99.91%	99.56%
100%	1601711	10	9.98	99.18%	
150%	1479921	15	15.02	99.60%	

Table 10: Showing accuracy results for Cilastatin

%Concentration (at specification level)	Average area	Amount Added (mg)	Amount found (mg)	% Recovery	Mean recovery
50%	1656163	0.5	0.99	99.53%	99.47%
100%	1920586	1.0	1.05	99.38%	
150%	2058167	1.5	1.495	99.52%	

5. References

1. AbuRuz, S., Millership J, McElnay J. The development and validation of liquid chromatography method for the simultaneous determination of metformin and glipizide, gliclazide, glibenclamide or glimiperide in plasma. *J Chromatogr. B*, **2005**, 817(2): 277-286.
2. Alexandar S, Diwedi R, Chandrasekar M. A RP-HPLC method for simultaneous estimation of metformin and pioglitazone in pharmaceutical formulation. *Res. J. Pharm. BioChemica. Sci.*, **2010**, 1(4): 858-866.
3. Al-Rimawi F, Development and validation of an analytical method for metformin hydrochloride and its related compound (1-cyanoguanidine) in tablet formulations by HPLC-UV. *Talanta*. **2009**; 79(5): 1368-71.
4. Aurelian E.Ranetti, Camelia Elena Stecoza, Constantin Mircioiu. Validation of a HPLC method for the simultaneous analysis of metformin and gliclazide in human plasma, *Farmacia*. **2009**, 57 (6): 729-735.
5. Bala Sekaran C, Prameela Rani A. Development and Validation of spectrophotometric method for the determination of DPP-4 Inhibitor, Sitagliptin, in its pharmaceutical preparations. *Int. J. Pharm. Pharm. Sci.*, **2010**, 2(4): 138-142.
6. Campbell DB, Lavielle R, Nathan C. The mode of action and clinical pharmacology of gliclazide: a review. *Diab Res Clin Prac.*, **1991**, 14: S21-S36.
7. Dubal Anil, Khatwal Rizwanbasha, Kosaraju Jayasankar, Meda Venkat, Samanta Malay. Bioanalytical method development and validation of sitagliptin phosphate by RP-HPLC and its application to pharmacokinetic study. *Int. J. Pharm Pharm Sci.*, **2012**, 4(2): 691-694.
8. Florentin T, Monica A .Specificity of an analytical hplc assay method of metformin hydrochloride. *Revue Roumaine de Chimie*.2007; 52(6):603–609.

9. Freddy H. Havaladar, Dharmendra L.Vairal. Simultaneous estimation of metformin hydrochloride, rosiglitazone and pioglitazone hydrochloride in the tablets dosage form. *Int. J.Appl. Bio. Pharm.Tec.*, **2010**, 1(3): 1000-1005.
10. Georgita C, Albu F, David V, Medvedovici A. Simultaneous assay of metformin and glibenclamide in human plasma based on extraction-less sample preparation procedure and LC/(APCI)MS. *J. Chromatogr. B.* **2007**, 854(1-2): 211-218.
11. Ghazala Khan, Dinesh SahuAgrawal Y P, NeetuSabarwal, AvnishJain,Gupta A K. Simultaneous Estimation of Metformin and Sitagliptin In Tablet Dosage Form. *Asian J. Biochem. Pharma. Res.*, **2011**; 1(2): 352-358.
12. Hassa Saad S.M., Mahmoud Wagiha H., Elmosallamy Mohamed A.F, Othman Abdel Hammeed M. Determination of metformin in pharmaceutical preparations using potentiometry, spectrofluorimetry and UV-visible spectrophotometry. *Anal. Chimica Acta.* **1999**; 378(1-3): 299-311.
13. Havele S, Dhaneshwar S. Development and validation of a HPLC method for the determination of metformin hydrochloride, gliclazide and pioglitazone hydrochloride in multicomponent formulation. *Webmed central pharmaceutical sciences.* **2010**, 1(10)
14. Havele S, Dhaneshwar S. Estimation of Metformin in Bulk Drug and in Formulation by HPTLC. *J Nanomedic Nanotechnolo.* **2010**, 1: 102. doi:10.4172/2157-7439.1000102
15. Herman G, Bergman A, Liu F, Stevens C, Wang A, Zeng W, Chen L, Snyder K, Hilliard D, Tanen M, Tanaka W, Meehan A, Lasseter K, Dilzer S, Blum R, Wagner J. Pharmacokinetics and pharmacodynamic effects of the oral DPP-4 inhibitor sitagliptin in middle-aged obese subjects. *J.Clin.Pharmacol.* **2006**, 46(8): 876–886.
16. Lakshmi KS, Rajesh T, Sharma S, Lakshmi S. Development and Validation of Liquid Chromatographic and UV Derivative Spectrophotometric Methods for the Determination of Metformin, Pioglitazone and Glimepiride in Pharmaceutical Formulations. *Der Pharma Chemica.* **2009**, 1 (1): 238-246.
17. Lakshmi KS, Rajesh T, Sharma S. Simultaneous determination of metformin and pioglitazone by reversed phase HPLC in pharmaceutical dosage forms. *Int.J Pharm. Pharm. Sci.* 2009; 1(2): 162-166.
18. Maria-Cristina Ranetti, Mihaela Ionescu, Lavinia Hinescu, Elena Ionic, Valentina Anu a, Parag Pathade, Imran Md, Vinod Bairagi, Yogesh Ahire. Development and Validation of Stability Indicating UV Spectrophotometric Method for the Estimation of Sitagliptin Phosphate in Bulk and Tablet Dosage Form. *J. Pharm. Res.*, **2011**; 4(3): 871-873.
19. Patil S.S., Bonde C. G. Development and Validation of analytical method for Simultaneous Estimation of Glibenclamide and Metformin HCl in Bulk and Tablets using UV visible spectroscopy. *Int. J. ChemTec. Res.*, **2009**, 1(4): 905-909.
20. Pawar S, Meshram G, Jadhav R, Bansal Y. Simultaneous determination of Glimepiride and Metformin hydrochloride impurities in sustained release pharmaceutical drug product by HPLC. *Der Pharma Chemica.* **2010**, 2(4): 157-168.
21. Ramakrishna Nirogi, Vishwottam Kandikere, Koteswara Mudigonda, Prashanth Komarneni, Raghupathi Al eti, Rajeshkumar Boggavarapu. Sensitive liquid chromatography tandem mass spectrometry method for the quantification of sitagliptin, a DPP-4 inhibitor, in human plasma using liquid-liquid extraction. *Biomed. Chromatogr.* **2008**, 22(2): 214–222.
22. Ramzia El-Bagary I, Ehab Elkady F, Bassam Ayoub M. Spectrofluorometric and Spectrophotometric Methods for the Determination of Sitagliptin in Binary Mixture with Metformin and Ternary Mixture with Metformin and Sitagliptin Alkaline Degradation Product. *Int. J. Biomed. Sci.*, **2011**, 7(1): 62-69.
23. Ravi Pratap Pulla, Sastry B S, Rajendra Prasad Y, AppalaRaju N. Simultaneous Estimation of Metformin HCl and Sitagliptin Phosphate in Tablet Dosage Forms by RP-HPLC. *Res. J. Pharm. Tech.* **2011**, 4(4): 646-649.
24. Robert Moses, Fixed combination of repaglinide and metformin in the management of type 2 diabetes, *Diabetes, Metabolic syndrome and obesity: Targets and Therapy* Dove press, open access to scientific and medical research. **2009**, 2: 101-9.
25. Shyamala M, Mohideen S, Satyanarayana T, Narasimha Raju Ch, Suresh Kumar P, Swetha K. Validated RP-HPLC for simultaneous estimation of Sitagliptin phosphate and Metformin in hydrochloride in tablet dosage form. *American J. Pharm Tech. Res.*, **2011**, 1(2): 93-101.
26. Tripathi K.D. *Essential of Medical Pharmacology*, 5th Edn, Jaypee Brothers Medical publisher New Delhi. pp: 515-516.
27. Wei Zeng, Donald Musson G, Alison Fisher L, Li Chen, Michael Schwartz S, Eric Woolf J, Amy Qiu Wang. Determination of sitagliptin in human urine and hemodialysate using turbulent flow online extraction and tandem mass spectrometry. *J. Pharm. Biomed. Anal.* **2008**, 46(3): 534-542.
28. Zeng W, Xu Y, Constanzer M, Woolf EJ. Determination of sitagliptin in human plasma using protein precipitation and tandem mass spectrometry. *J. Chromatogr. B.* **2010**, 878(21): 1817-1823.