

International Journal of Current Trends in Pharmaceutical Research



Journal Home Page: www.pharmaresearchlibrary.com/ijctpr

RESEARCH ARTICLE

Assessment of Antimicrobial Activity of Aniline and Pyridine Derivatives by Cup Plate Method

Aiswarya Moorthy^{*1}, Cholaraja Kamatchi², Jeyaraman Amutha Iswarya Devi³

Department of Pharmaceutical Chemistry, Arulmigu Kalasalingam College of Pharmacy, Anand Nagar, Krishnankoil-626126, Srivilliputtur (via) Tamil Nadu, India

ABSTRACT

The discovery of antibiotics, and the subsequent development of synthetic antimicrobial compounds, altered our therapeutic approach towards infectious diseases, and improved the quality and length of life for humans and other organisms. In present study investigate the antimicrobial properties of aniline and pyridine derivatives (SAHA and MS-275). The compound were synthesized in laboratory, Muller Hinton agar media and Sabouraud dextrose agar media was used to investigate the antibacterial and antifungal activity respectively, both activities followed by gel diffusion principle in cup plate method. **Keywords:**Antimicrobial properties, Cup plate method, Gel diffusion, SAHA and MS-275.

ARTICLE INFO

Corresponding Author Aiswarya Moorthy Department of Pharmaceutical Chemistry, Arulmigu Kalasalingam College of Pharmacy, Krishnankoil, Tamil Nadu, India MS-ID: IJCTPR3939



PAPER QR-CODE

Article History: Received 19 Jan 2019, Accepted 27 February 2019, Available Online 15 May 2019

Copyright© 2019 Aiswarya Moorthy, et al. Production and hosting by Pharma Research Library. All rights reserved.

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited.

Citation: Aiswarya Moorthy, et al. Assessment of Antimicrobial Activity of Aniline and Pyridine Derivatives by Cup Plate Method. Int. J. Curnt. Tren. Pharm, Res., Res., 2019, 7(3): 91-94.

CONTENTS

1. Introduction 2. Materials and Methods	
3. Results and Discussion	93
4. Conclusion	94
5. References	94

1. Introduction

The science dealing with the study of the prevention and treatment of diseases caused by micro-organisms is known as medical microbiology. The detection and development of antibiotics and the consequent consumption of antibiotic therapy signaled a milestone in the history of biology and International Journal of Current Trends in Pharmaceutical Research

medicine. Antibiotic therapy permitted for the treatment of dangerous and fatal diseases, leading to a remarkable development in health conditions for humans, as well as rising life expectancy. In addition, antibiotic therapy provided significant health benefits for animals and

Aiswarya Moorthy et al, IJCTPR, 2019, 7(3): 91-94

vegetation, improving the production and consumption of reared organisms and human food crops. For the treatment of ailment inhibitory chemicals working to kill microorganisms or stop their growth, are called antimicrobial agents [1,2]. These are categorized according to their use and spectrum of activity, as germicides that kill microorganisms, whereas micro-biostatic agents inhibit the growth of pathogens and enable the leucocytes and other defense mechanism of the host to cope up with static invaders [3]. The germicides may exhibit selective toxicity depending on their spectrum of activity. They may act as viricides (killing viruses), bacteriocides (killing bacteria), algicides (killing algae) or fungicides (killing fungi). Paul Ehrlich used the word chemotherapy for curing the communicable disease without damage to the host's tissue, known as chemotherapeutic agents such as antibacterial, antiprotozoal, antiviral, antineoplastic, antitubercular and antifungal agents [4,5]. A few of the early antibiotics are at the present known to be unacceptably toxic and are therefore not used in antimicrobial therapy. Some however, such as adriamycin, bleomycin and mitomycin are presently used as cytotoxic agents in cancer therapy [6].



Fig 1:Phenyl amide Octanedioic acid hydroxyamide (SAHA)



Fig 2: pyridine-3-ylmethyl{4-[aminophenyl)carbamoyl]benzyl}carbamate (MS-275)

2. Materials and Methods Materials:

The test organisms include the following bacterisa and likes Staphylococcus aureus, Pseudomonas fungi aeruginosa and Candida albicans, all the strains were obtained from Institute of Microbial technology, Chandigarh, India; Sabouraud dextrose agar media and Muller Hinton agar media(Himedia, Mumbai); Aniline, DMF, THF were obtained from Sisco Research Lab Pvt. Ltd., New Mumbai; Suberic acid, EDCI, HOBt.H2O, N,Ndisopropyl ethylamine, Pyridine-3-methanol, CDI, were obtained from Sigma-Aldrich, Missouri, U.S; 4-amino methyl benzoic acid, DBU, Triethylamine, were obtained from TOKYO chemicals Pvt. Ltd, Chennai, India. All the Chemicals and reagents used in this experiment are of analytical grade. Both the compounds (SAHA and MS-275) are aniline and pyridine derivatives were synthesized in our laboratory [7-10]. In this experiment the synthesized compounds are used for anti-bacterial and anti-fungal activities.

Antibacterial Activity of Synthesized Compounds Organism used - Staphylococcus aureus [Gm (+Ve)], International Journal of Current Trends in Pharmaceutical Research

Pseudomonas aeruginosa[Gm (-Ve)]			
Standard used	- A	mikacin	
Incubation temperature -37°C			
Incubation time	-	24 hrs	
Control	-	CH ₃ OH	
Test concentratio	n -	50 mcg / ml	
Method	-	Cup- plate method	
Principle	-	Gel diffusion	
Composition of Muller Hinton agar media:			
Beef extract	-	30 g	
Peptone	-	17.5 g	
Starch	-	1.5 g	
Agar	-	17 g	
NaOH	-	5 g	
Distilled water	-	1000 ml	
Final pH at 25° C-7.4 ± 0.2			

Method

The beef extract, peptone, starch and agar were taken in the above proportions and dissolved in 100 ml of distilled water. The constituents were heated gently at 100°C with agitation. The pH of medium was adjusted to 7.4 using sodium hydroxide. It is transferred to boiling tube in hot condition and sealed with non-absorbent cotton and sterilized by autoclaving at 121°C (15 lbs pressure) for 15 minutes, then poured aseptically into sterile Petri dishes. Aseptic condition is maintained throughout the process.

Inoculation of microorganisms:

For the screening of antibacterial activity Cup- plate method was used. The sterilized Muller Hinton Agar media was heated on a water bath to melt the media. When media was lukewarm, the organisms were inoculated separately and poured aseptically into sterile Petri dishes and allowed to solidify. The temperature of the medium should not exceed about 50°C when organisms were inoculated. The solutions of known concentration standard drug Amikacin (50µg/ml) and corresponding concentration of synthesized compounds (50µg/ml) were prepared. Apply the solutions to the surface of the medium in sterile cavities prepared in the media. One more solution of Methanol is used as control in separate cavity. The volume of solution added to each cavity must be uniform and sufficient to fill the hole. Petri dishes were incubated at 37°C for 24 hours, after placing them in refrigerator for one hour to facilitate uniform diffusion. Observations were made for the zone of inhibition around the synthesized compounds and compared with that of standard.



Fig 3: Zone of inhibition against S.aureus



Fig 4: Zone of inhibition against Pseudomonas aeruginosa

Antifungal Activity

Organisms used	-	Candida albicans
Standard used	-	Ketoconazole (10 µg / ml)
Incubation temperature	-	28°C
Incubation time	-	24 hrs
Control used	-	CH ₃ OH
Test concentration	-	5 µg / ml
Method	-	cup plate method
Principle	-	gel diffusion
Composition of Sabour	aud	dextrose agar media:
Cluses 40 cm		-

Glucose-	40 gm
Peptone -	10 g
Agar -	15 g
Distilled water	- 1000 ml
Final Ph-	5.4
Method:	

Glucose, peptone and agar were taken in the above proportions and dissolved up to 1000 ml of distilled water. The constituents were heated gently at 100°C with agitation. The pH of media was adjusted to 5.4. Then it was transferred to a boiling tube in hot condition and sealed with non-absorbent cotton and sterilized by autoclaving at 121°C (15 lbs pressure) for 15 mins, then poured aseptically into sterile Petri dish.

Inoculation of microorganism:

For the screening of antifungal activity disc Cup-plate method was used. Sabouraud dextrose agar plates were allowed to solidify and inverted to prevent the condensate falling on the agar surface. The plates were dried at 37°C just before inoculation agar plates were incubated for about 24 hours at 37°C, which showed sufficient growth of fungi. The temperature of the medium should not exceed about 50°C when organisms were inoculated.

The solutions of known concentration standard drug Ketoconazole (10µg/ml) and corresponding concentration of synthesized compounds (10µg/ml) were prepared. Apply the solutions to the surface of the medium in sterile cavities prepared in the media. One more solution of Methanol is used as control in separate cavity. The volume of solution added to each cavity must be uniform and sufficient to fill the hole. Petri dishes were incubated at 37°C for 24 hours, after placing them in refrigerator for one hour to facilitate uniform diffusion. Observations were made for zone of inhibition around the synthesized compounds with that of standard.

CODEN (USA): IJCTGM | ISSN: 2321-3760



Fig 5: Zone of inhibition against Candida albicans

3. Results and Discussion

In this present study investigated the antimicrobial activity of synthesized aniline and pyridine derivatives (SAHA and MS-275). The zone of inhibition was measured in cup plate method. According to the table: 1 the synthesized aniline derivative was potent against gram positive bacteria; pyridine derivative was potent against gram negative bacteria and its compare with standard drug. The both compounds SAHA and MS-275 also have anti-fungal activity. In this experiment the zone of inhibition was measured and reported in table: 2.

Fable	1:Zone	of i	inhib	ition
-------	--------	------	-------	-------

	Zone of inhibition against			
Compounds	Gram (+)Ve	Gram(-)Ve in		
	S.aureus in mm	P. aeruginosa in mm		
SAHA	11	13		
MS-275	14	17		
Control	Desistant	Desistant		
(Methanol)	Resistant	Resistant		
Standard	17	10		
(Amikacin)	17	18		

Table 2: Zone of inhibition

Compounds	Zone of inhibition in
	mm by Candida albicans
SAHA	19
MS-275	18
Control (Methanol)	Resistant
Standard (Ketoconazole)	16



Fig 6: Antibacterial activity

International Journal of Current Trends in Pharmaceutical Research

Aiswarya Moorthy et al, IJCTPR, 2019, 7(3): 91-94



Fig 7: Anti-fungal activity

4. Conclusion

Continuous and recurrent use of antibiotics to manage human illness has caused by bacteria to obtain resistance against the frequently used therapeutic agents. As a result the difficulties of microbial resistance is rising and require to look for alternative newer molecules is increasing. The present investigation has confirmed the synthesized aniline and pyridine derivatives have potential antibacterial and antifungal properties and they need further investigations for the therapeutic value in managing diseases caused by the test bacteria based on toxicological and pharmacological study.

5. References

- [1] Aminov, R.I. A brief history of the antibiotic era: Lessons learned and challenges for the future. Front. Microbiol. 1, 1–7, 2010.
- [2] Schwartz, M. Historical streptococci. In Streptococci and the Host; Horaud, T., Bouvet, A., Leclercq, R., de Montclos, H., Sicard, M., Eds.; Plenum Press: New York, NY, USA, Chapter 1, pp. 1–2 1997.
- [3] Fleming, A. On the antibacterial action of cultures of a Penicillium, with special reference to their use in the isolation of B. influenzae. Br. J. Exp. Pathol. 10, 226–236 1929.
- [4] Cohen ML. Epidemiology and drug resistance: implications for a post-antimicrobial era. Science.257: 1050, 1992.
- [5] Schatz, A.; Bugle, E.; Waksman, S.A. Streptomycin, a substance exhibiting antibiotic activity against gram-positive and gram-negative bacteria. Proc. Soc. Exp. Biol. Med., 55, 66– 69,1944.
- [6] Sköld, O. Antibiotics and Antibiotic Resistance; John Wiley & Sons, Inc.: Hoboken, NJ, USA, pp. 1–207,2011.
- [7] Willard H.H, instrumental methods of analysis, CBS publishers, 7th edition, 440-462 1986.
- [8] Bekette A.H and stenlake. J.B., Practical pharmaceutical chemistry, CBS publishers, 3rd edition, page 408 467, 1997.
- [9] Silverstein R.M., spectrometric identification of organic compounds, John Wiley and sons, 5th edition, page 102-131, 1991.

- [10] Skoog D.A., fundamentals of analytical chemistry, Harcouris asia, Pvt Ltd, , 7th edition, 592 – 597, 2001.
- [11] Alekshun, M.N.; Levy, S.B. Molecular mechanisms of antibacterial multidrug resistance. Cell, 128, 1037–1050, 2007.
- [12] Pacheco, R.; Correia, S.; Poeta, P.; Pinto, L.; Igrejas, G. The role of proteomics in elucidating multiple antibiotic resistance in Salmonella and in novel antibacterial discovery. In Salmonella— Distribution, Adaptation, Control Measures and Molecular Technologies; Annous, B.A., Gurtler, J.B., Eds.; InTech: Rijeka, Croatia, Chapter 10, pp. 187–220, 2012.
- [13] Newman, D.J.; Cragg, G.M.; Snader, K.M. Natural products as sources of new drugs over the period 1981–2002. J. Nat. Prod., 66, 1022–1037, 2003.
- [14] Franzenburg, S.;Walter, J.; Künzel, S.;Wang, J.; Baines, J.F.; Bosch, T.C.G.; Fraune, S. Distinct antimicrobial peptide expression determines host species-specific bacterial associations. Proc. Natl. Acad. Sci. USA, 110, E3730–E3738, 2013.
- [15] Harsh Mohan, text book of pathology, 5th edition, 133, 2005.
- [16] S.K. Kulkarni, hand book of experimental pharmacology, 3rd edition, 127, 1999.